



Inflammation in MPNs

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1. Employment

None

2. Advisory Board Activity

Pfizer, Incyte / Ariad, Novartis, AOP Pharma, BMS, Celgene, Geron, Janssen, CTI, Roche, Baxalta, Sanofi, MPN Hub, Sierra Oncology, Glaxo-Smith Kline, AbbVie, PharmaEssentia, MSD

3. Stock etc

None

4. Patents, Licences

RWTH Aachen University (Patent filed on own BET inhibitors)

5. Honoraria

Novartis, BMS, Pfizer, Incyte, Ariad, Shire, Roche, AOP Pharma, Janssen, Geron, Celgene, Karthos, Abbvie, iOMEDICO, MSD

6. Research Funding

Novartis Foundation, BMS, Novartis, AOP Pharma, Janssen/Geron

7. Other financial disclosures (e.g. travel support)

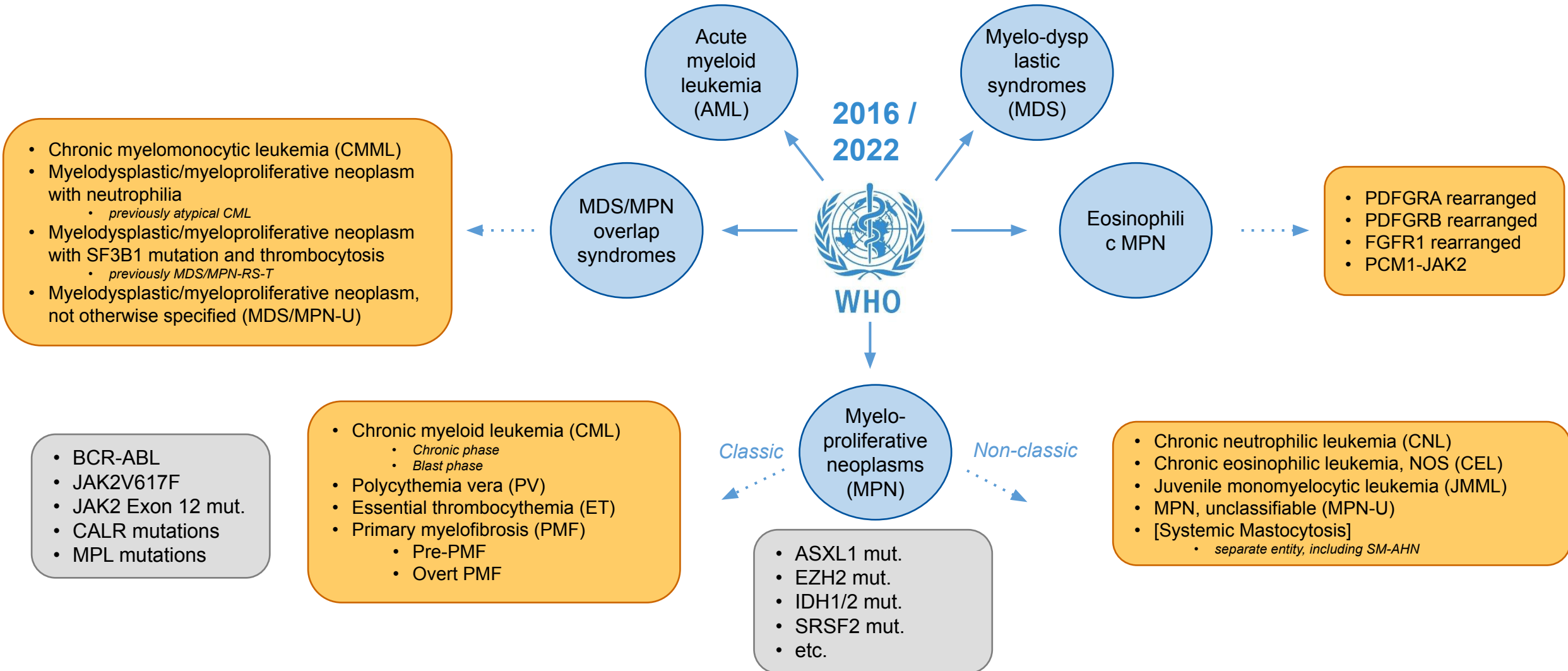
Alexion, Novartis, BMS, Incyte / Ariad, AOP Pharma, Baxalta, CTI, Pfizer, Sanofi, Celgene, Shire, Janssen, Geron, Karthos, Sierra Oncology, Glaxo-Smith Kline, Imago Biosciences, AbbVie, iOMEDICO, MSD

8. Immaterial disclosures

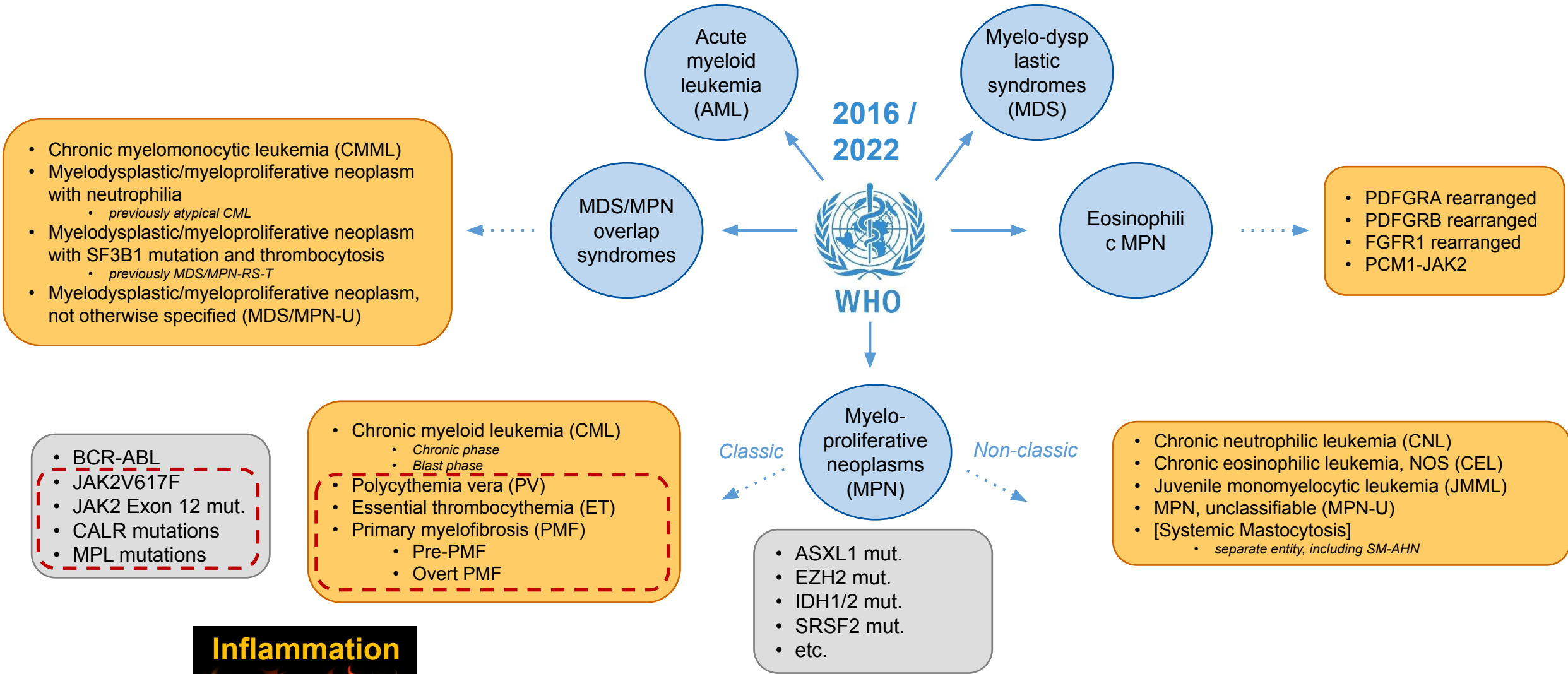
None

This presentation contains information on off-label therapies

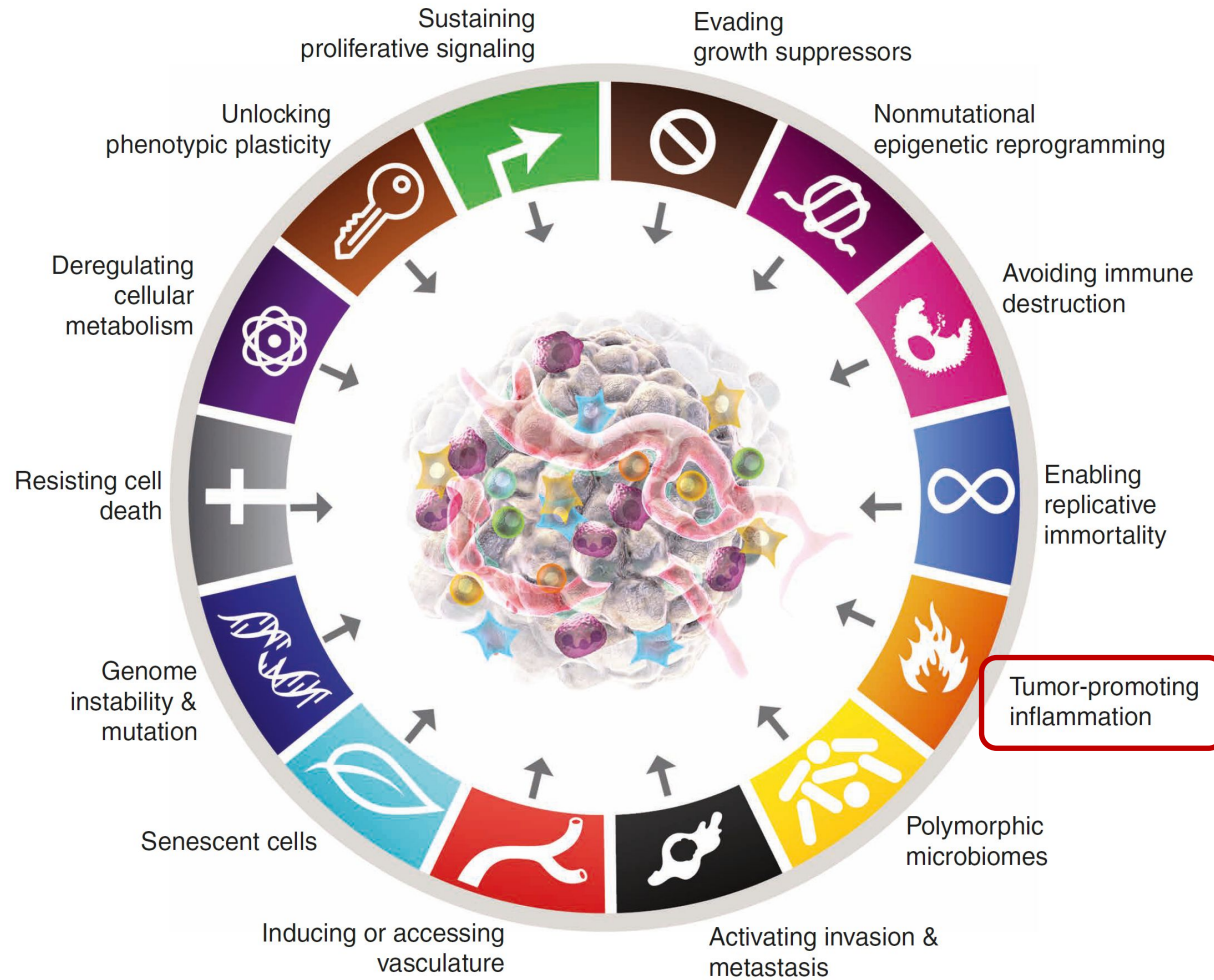
Myeloproliferative Neoplasms (MPN)



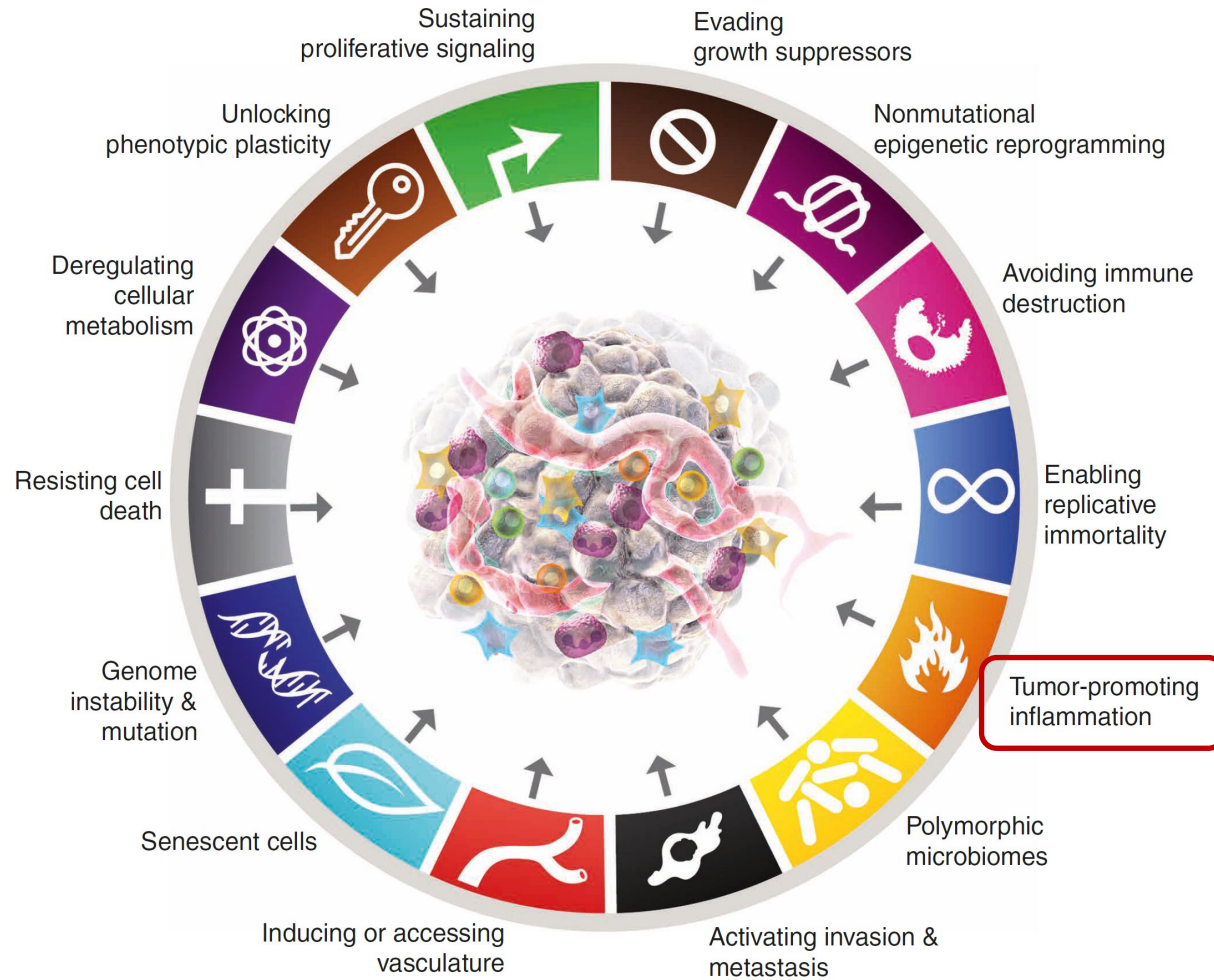
Myeloproliferative Neoplasms (MPN)



Inflammation and cancer: One of the hallmarks of cancer

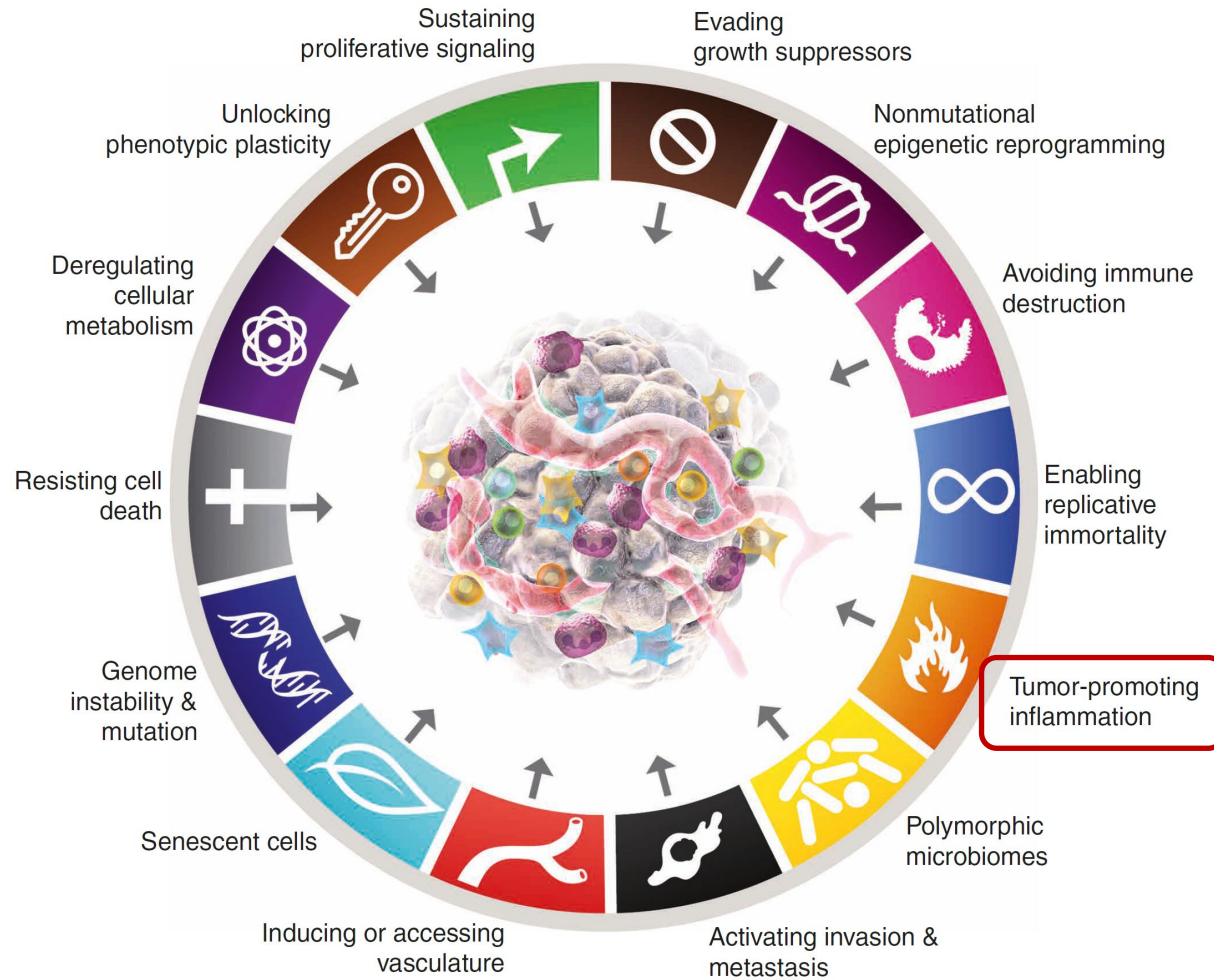


Inflammation and cancer: One of the hallmarks of cancer



**Myeloproliferative neoplasms (MPN)
are particular cancers...**

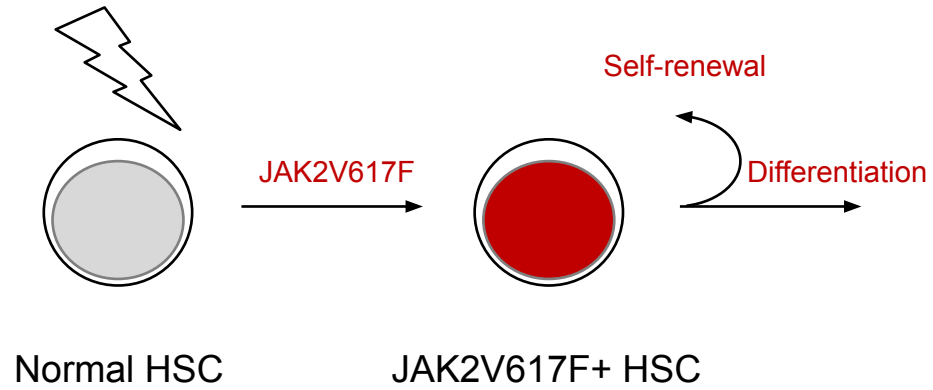
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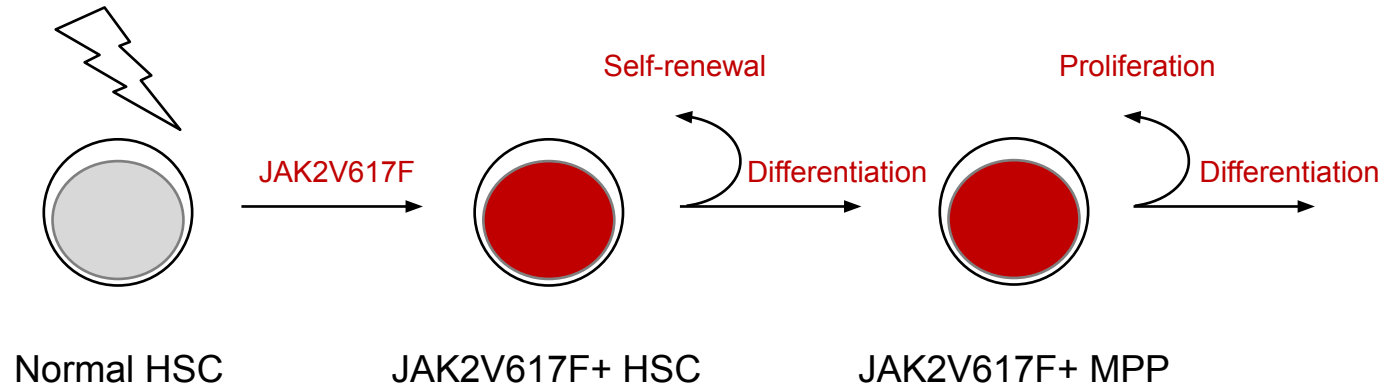
Myeloproliferative neoplasms (MPN) are particular cancers...

...since the clonal cancer cells themselves are inflammatory cells...

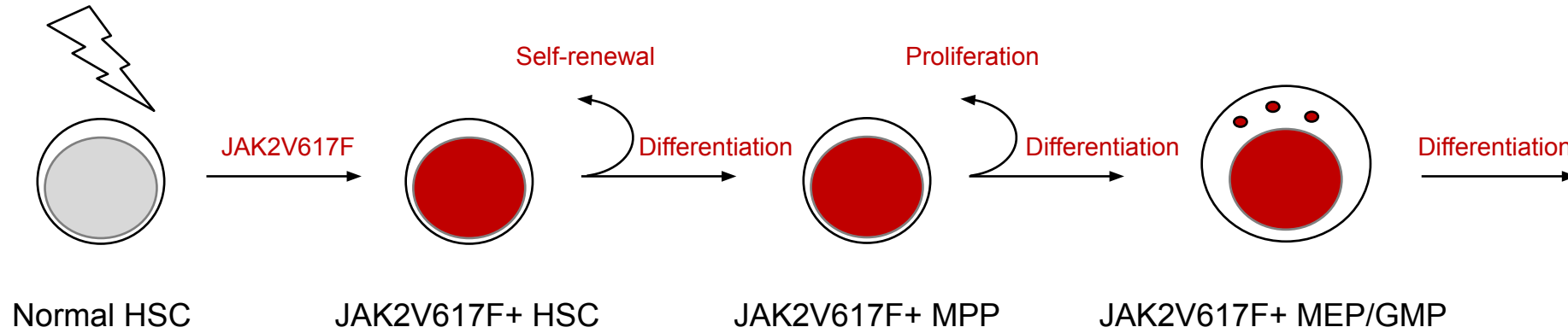
Personal view on MPN pathogenesis and the role of inflammation **UNIKLINIK RWTH AACHEN**



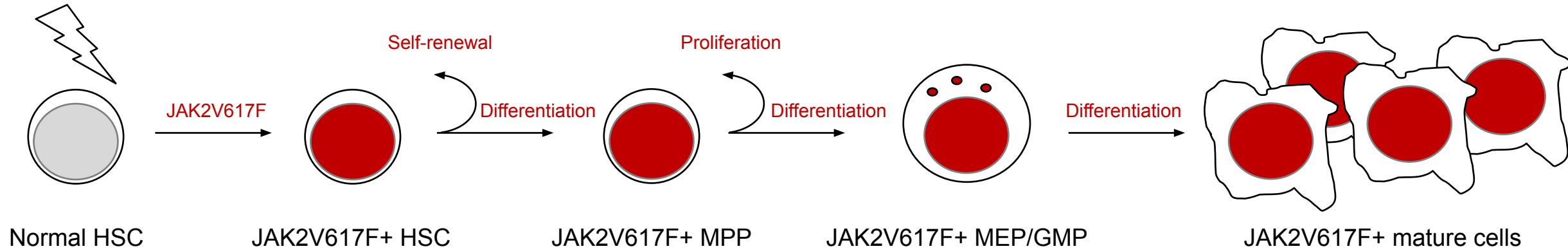
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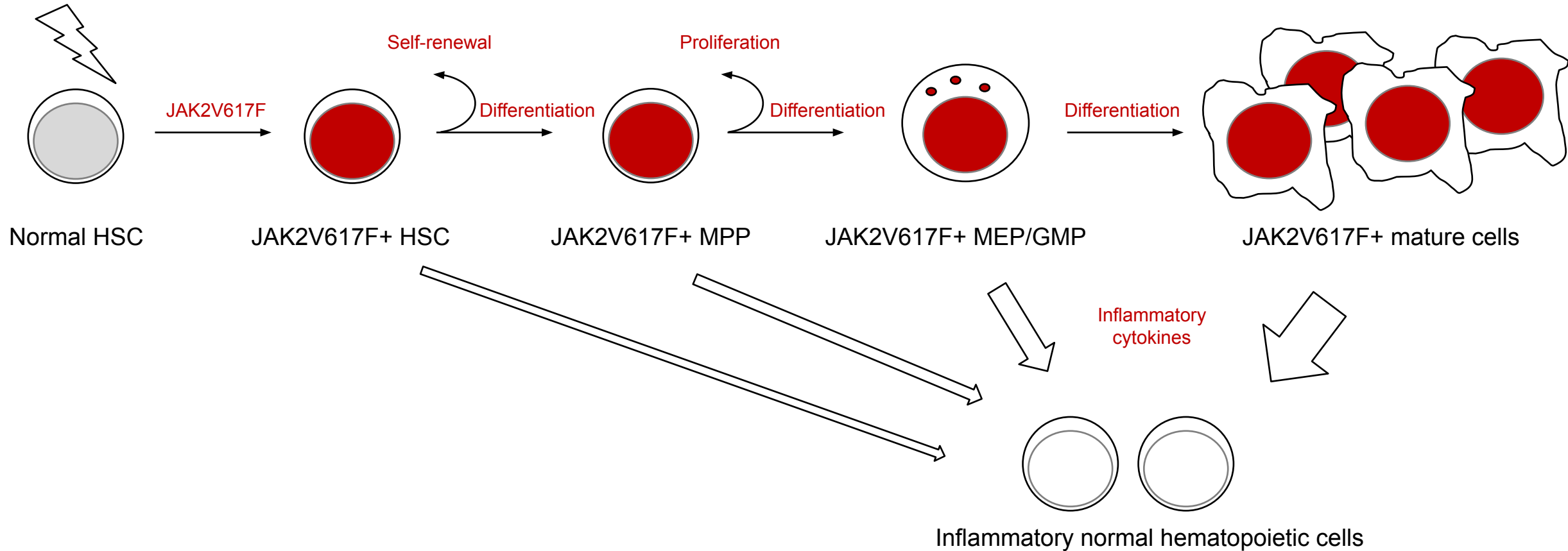
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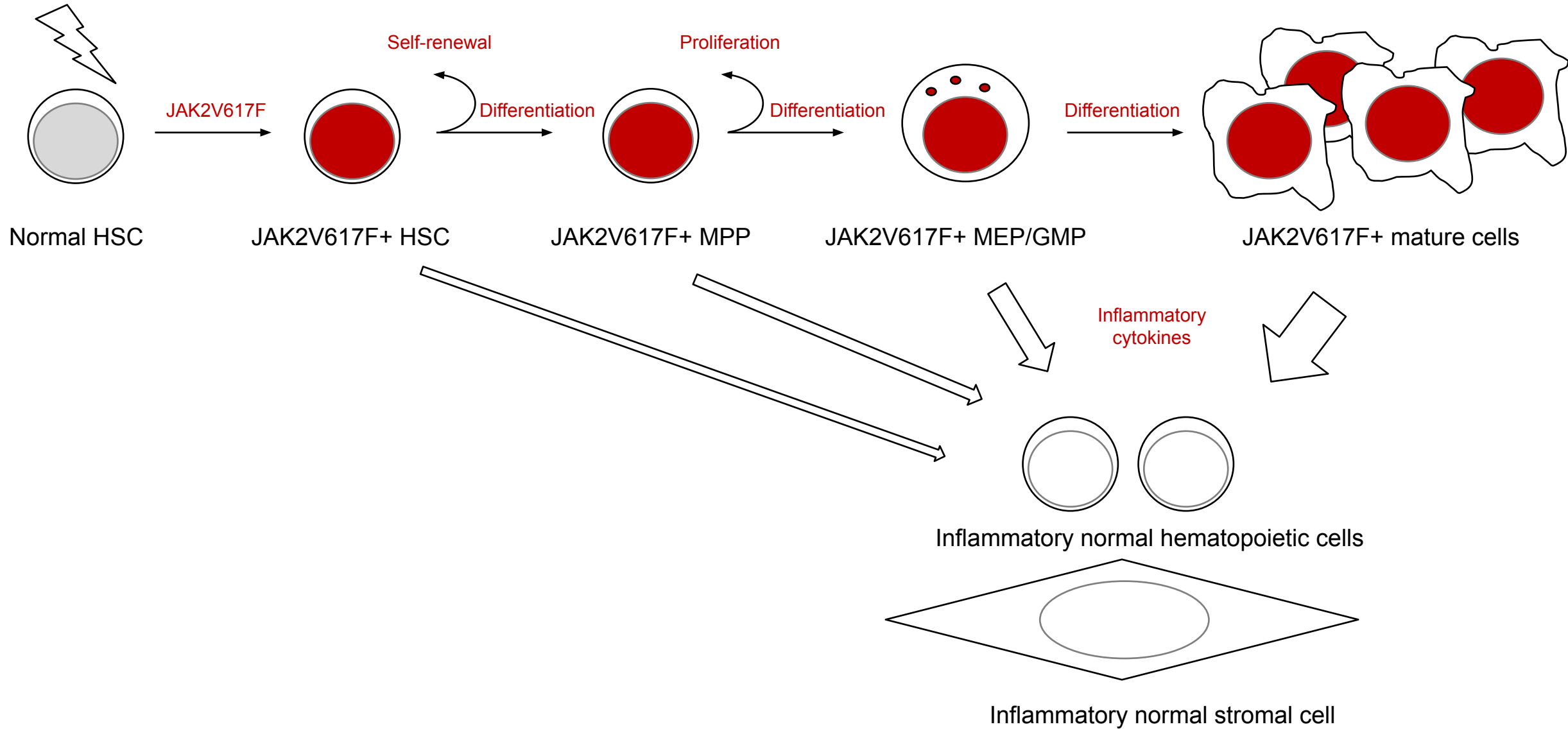
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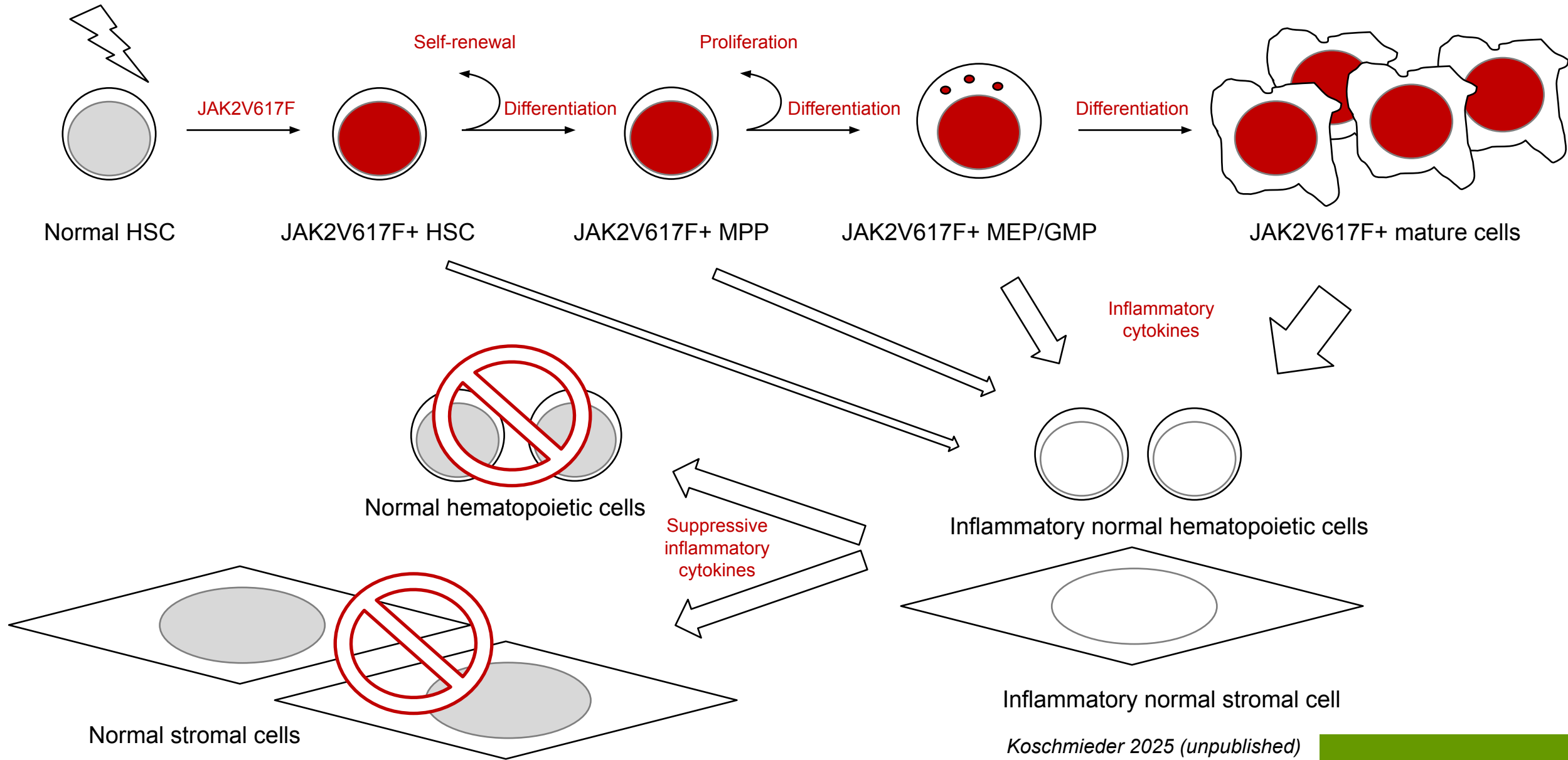
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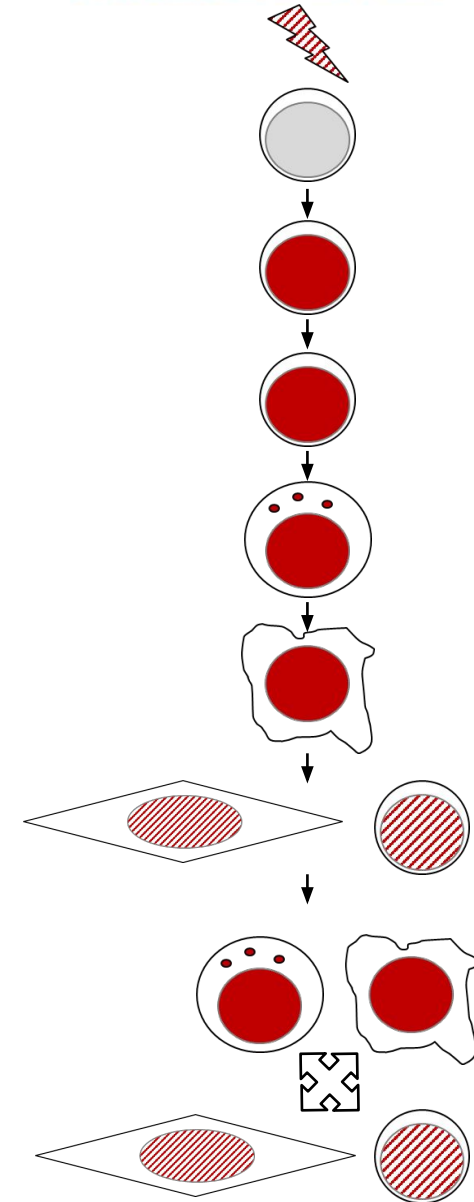


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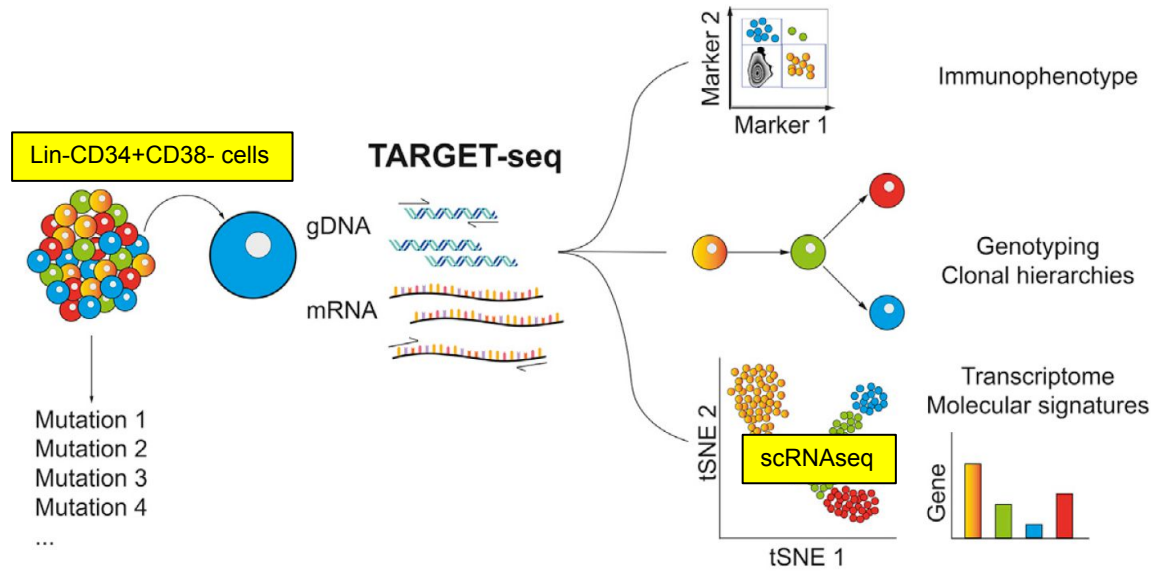


What is the evidence?

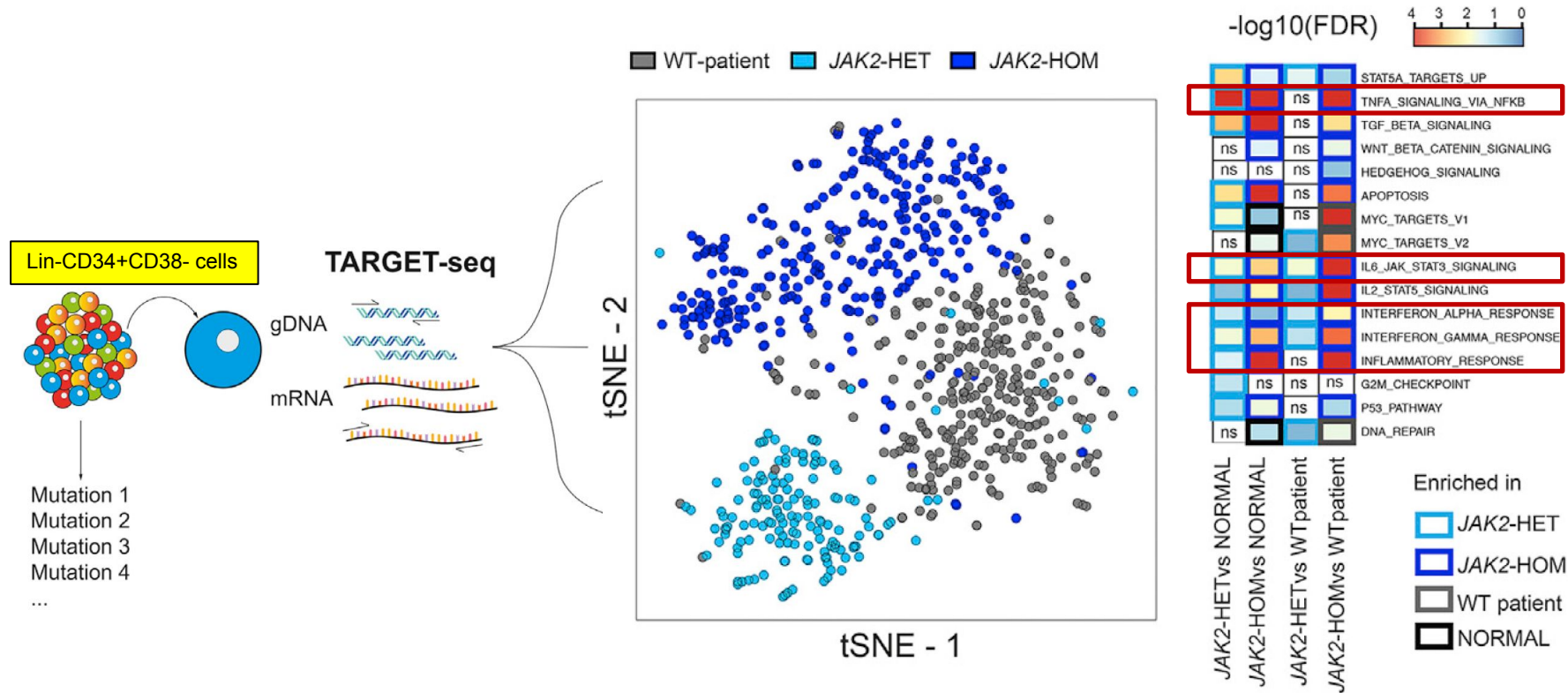
- HSC in the bone marrow (BM) acquires a somatic *JAK2V617F* (or CALR or MPL) driver mutation
 - This has been described to occur very early in life (e.g. in utero or early childhood)
 - *JAK2V617F* has been shown to induce cycling of HSCs



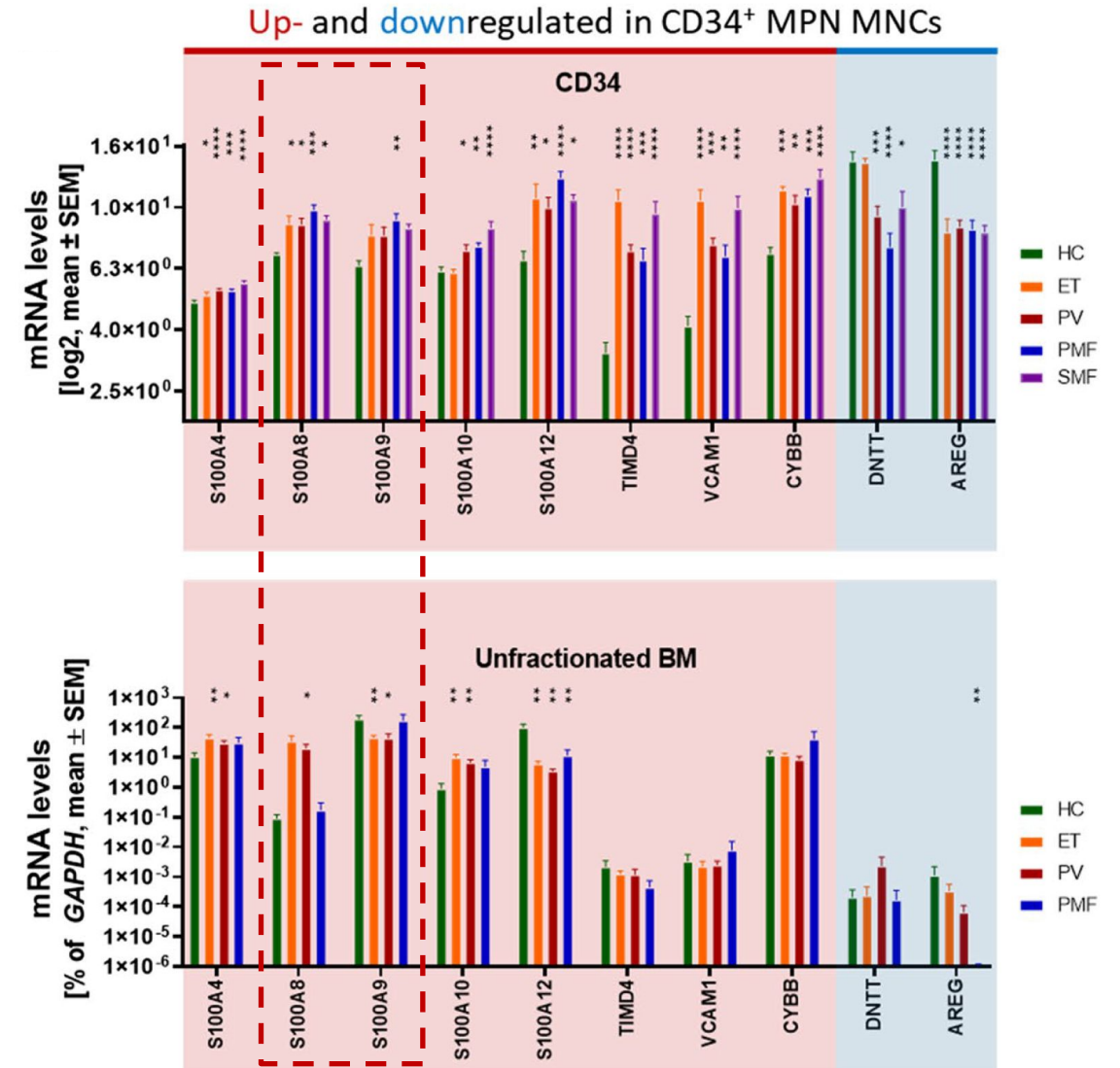
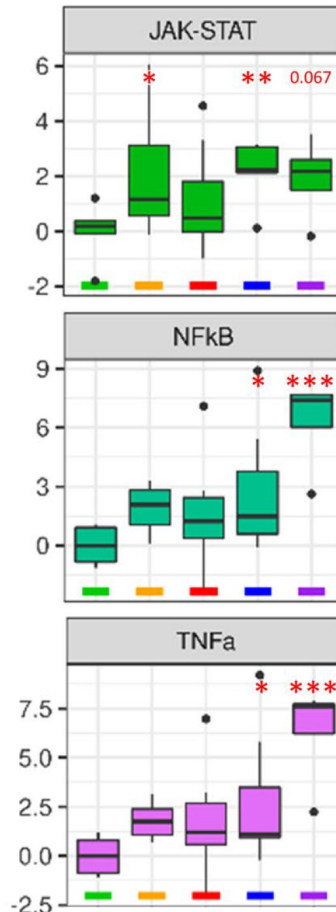
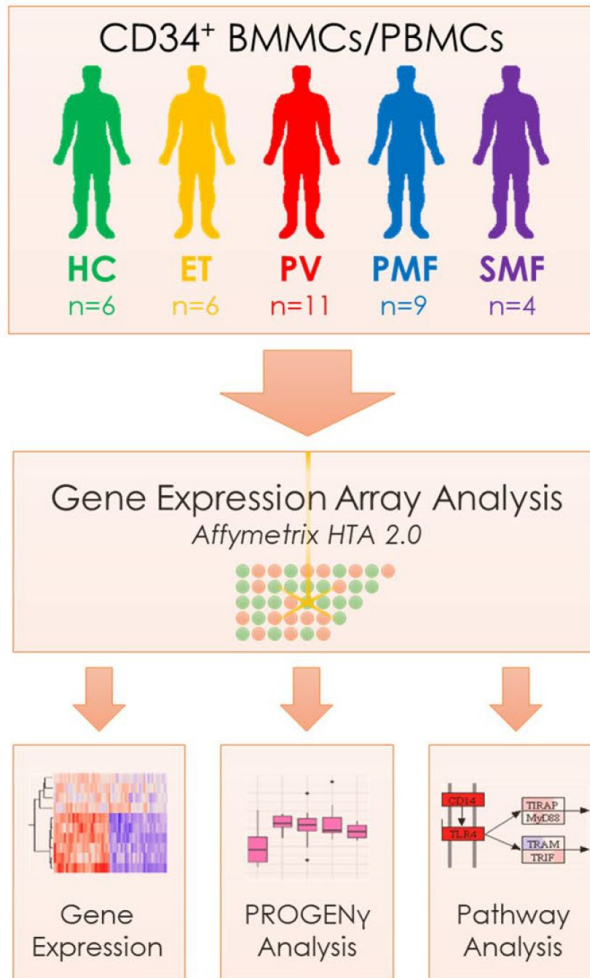
CD34+ cells from MPN patients express inflammatory genes



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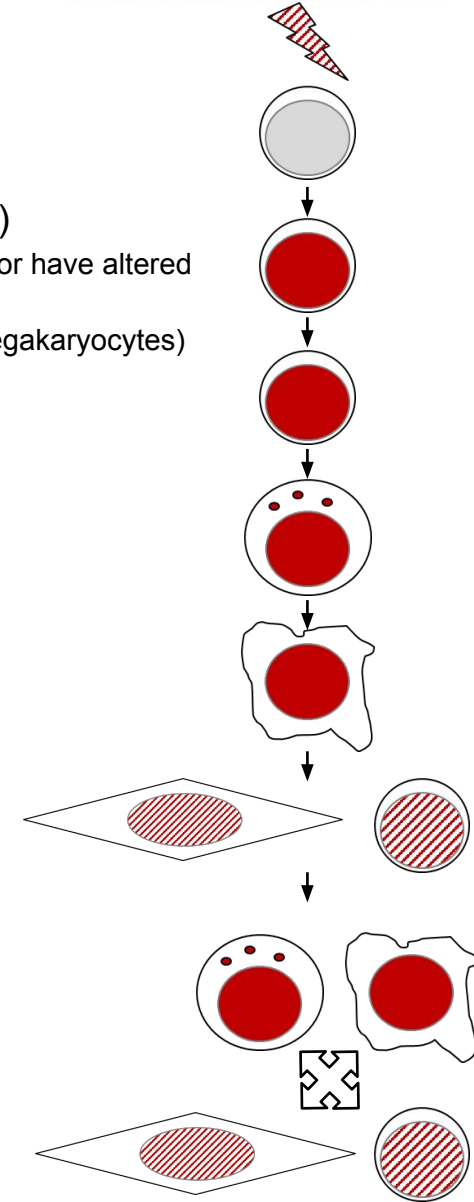


CD34+ cells from MPN patients express inflammatory genes



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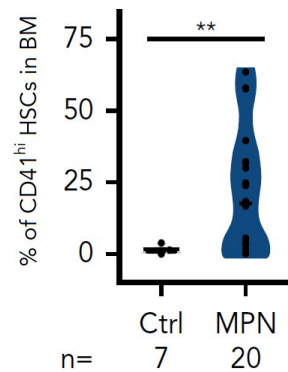
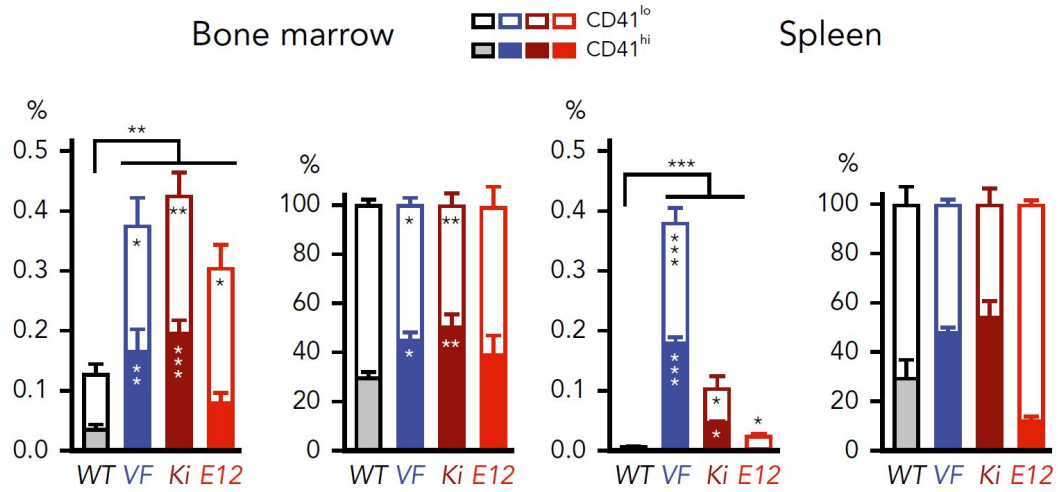
MPN-derived HSCs are skewed towards MKs

MYELOID NEOPLASIA

JAK2-V617F and interferon- α induce megakaryocyte-biased stem cells characterized by decreased long-term functionality

Tata Nageswara Rao,¹ Nils Hansen,¹ Jan Stetka,^{1,2} Damien Luque Paz,¹ Milena Kalmer,³ Julian Hilfiker,¹ Max Endeke,⁴ Nouraiz Ahmed,⁴ Lucia Kubovcakova,¹ Margareta Rybarikova,¹ Hui Hao-Shen,¹ Florian Geier,^{1,5} Christian Beisel,⁴ Stefan Dirnhofer,⁶ Timm Schroeder,⁴ Tim H. Brümmendorf,³ Dominik Wolf,⁷ Steffen Koschmieder,³ and Radek C. Skoda¹

Frequencies and percentages of CD41^{hi} and CD41^{lo} HSCs



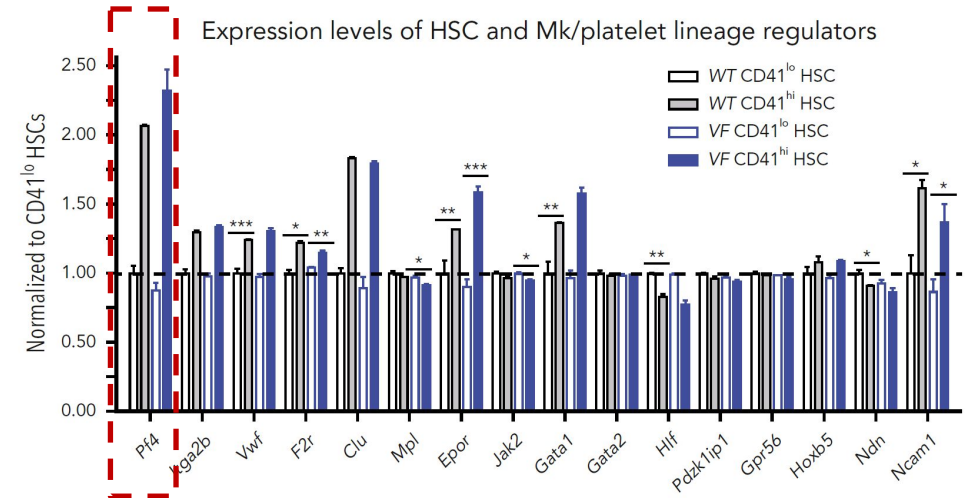
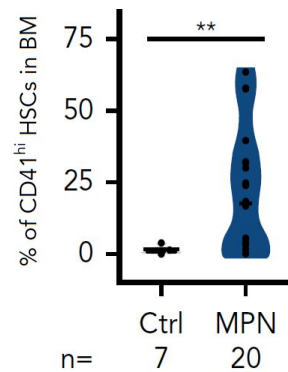
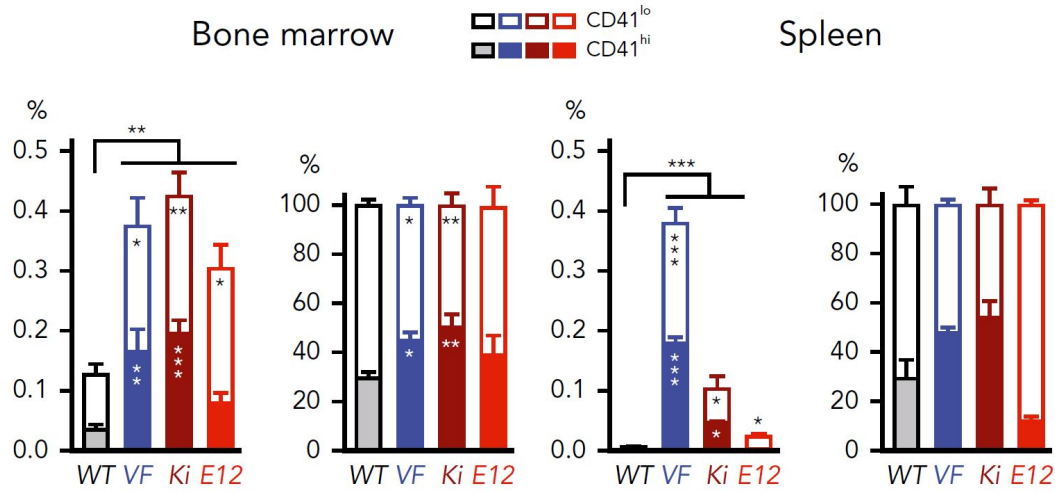
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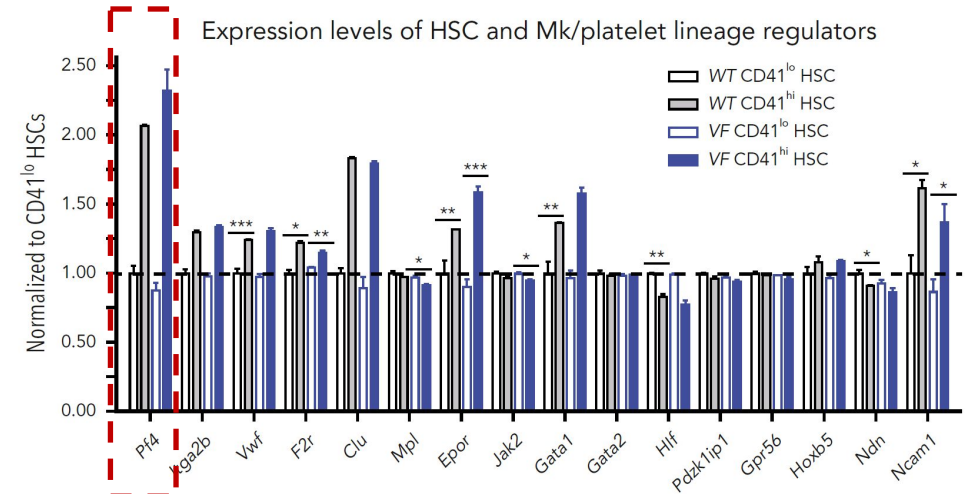
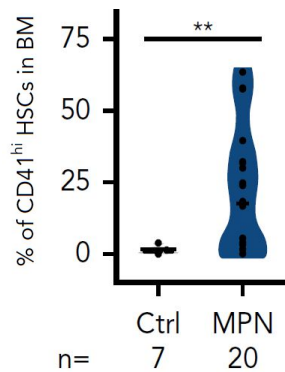
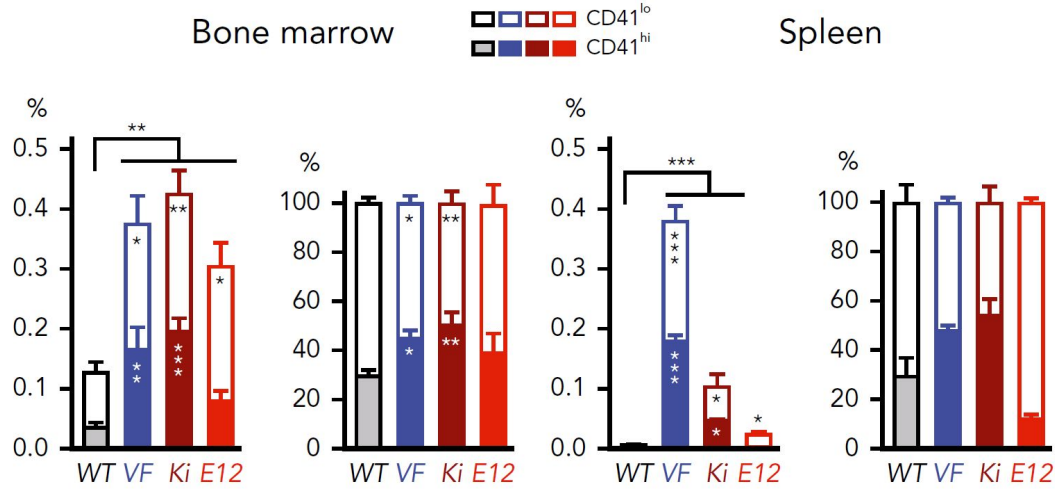
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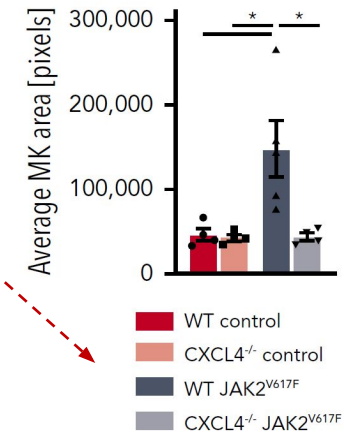
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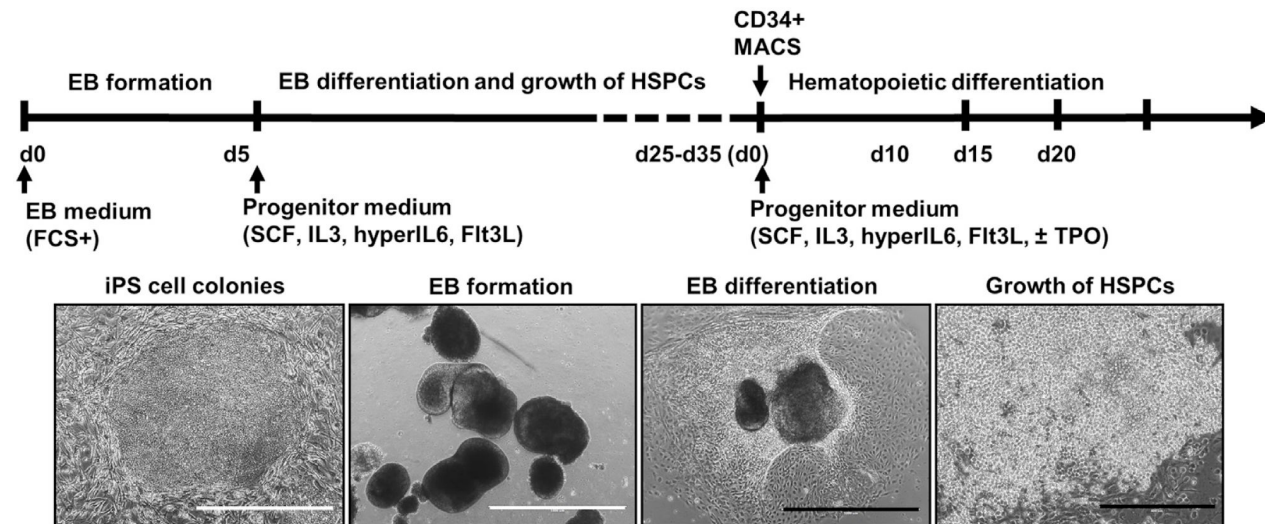
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Pf4/Cxcl4 axis



MPN-derived iPSCs are skewed towards MKs

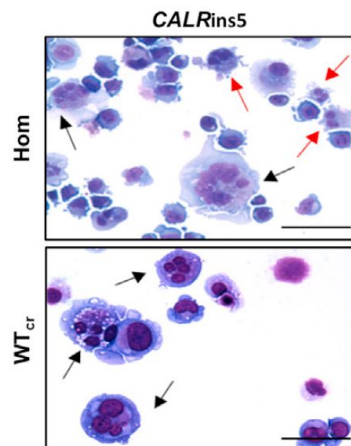
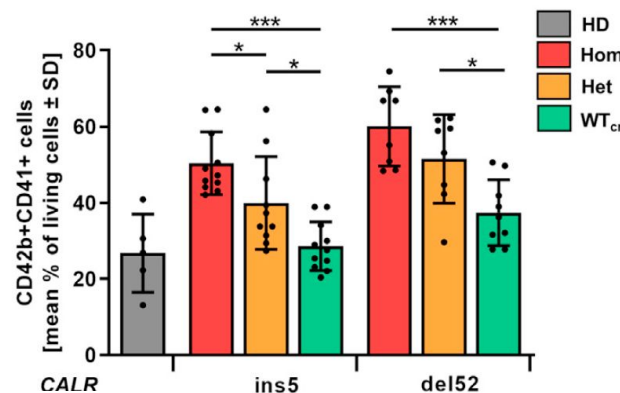


CALR frameshift mutations in MPN patient-derived iPSCs accelerate maturation of megakaryocytes

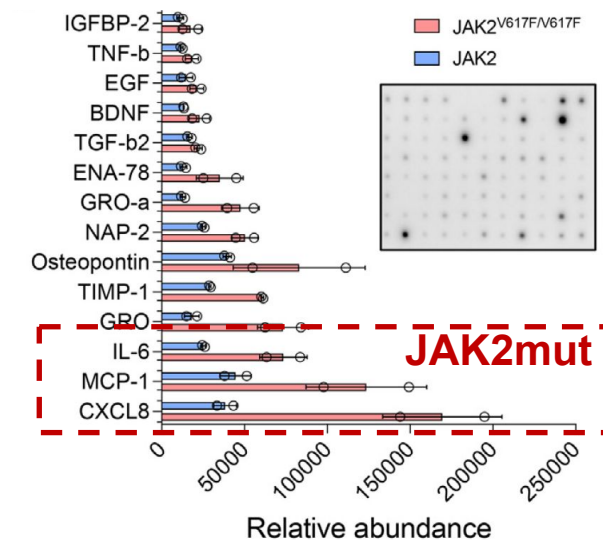
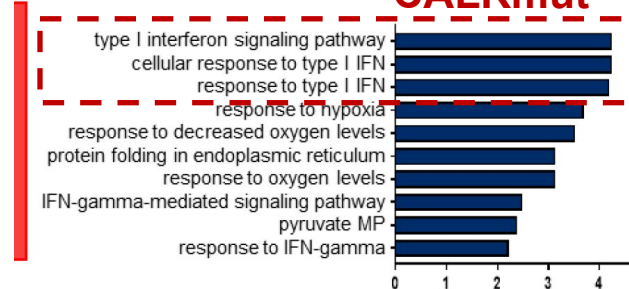
Kathrin Olschok,^{1,2,3,10} Lijuan Han,^{1,2,3,10} Marcelo A.S. de Toledo,^{1,2} Janik Böhnke,^{4,5} Martin Grafshoff,⁶ Ivan G. Costa,⁶ Alexandre Theocharides,⁷ Angela Maurer,^{1,2} Herdit M. Schüller,⁸ Eva Miriam Buhl,⁹ Kristina Pannen,^{1,2} Julian Baumeister,^{1,2} Milena Kalmer,^{1,2} Siddharth Gupta,^{1,2} Peter Boor,⁹ Deniz Gezer,^{1,2} Tim H. Brummendorf,^{1,2} Martin Zenke,^{4,5} Nicolas Chatain,^{1,2,11} and Steffen Koschmieder^{1,2,11,*}

Proinflammatory phenotype of iPS cell-derived JAK2 V617F megakaryocytes induces fibrosis in 3D *in vitro* bone marrow niche

Niclas Flösdorf,^{1,2,3,4} Janik Böhnke,^{1,2,4} Marcelo A.S. de Toledo,^{4,5} Niklas Lutterbach,³ Vanesa Gómez Lerma,^{1,2} Martin Grafshoff,⁶ Kathrin Olschok,^{4,5} Siddharth Gupta,^{4,5} Vithurithra Tharmapalan,^{2,4,7} Susanne Schmitz,³ Katrin Götz,³ Herdit M. Schüller,^{8,9} Angela Maurer,^{4,5} Stephanie Sontag,^{1,2} Caroline Küstermann,^{1,2} Kristin Seré,^{1,2,3} Wolfgang Wagner,^{2,4,7} Ivan G. Costa,⁶ Tim H. Brummendorf,^{4,5} Steffen Koschmieder,^{4,5} Nicolas Chatain,^{4,5} Miguel Castilho,¹⁰ Rebekka K. Schneider,³ and Martin Zenke^{1,2,4,5,*}

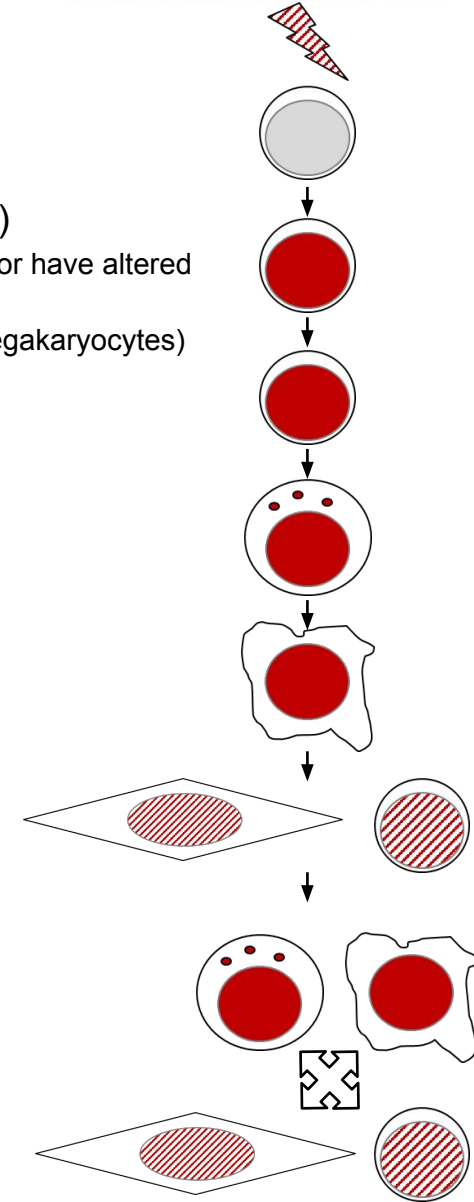


Upregulated GO terms CALRmut

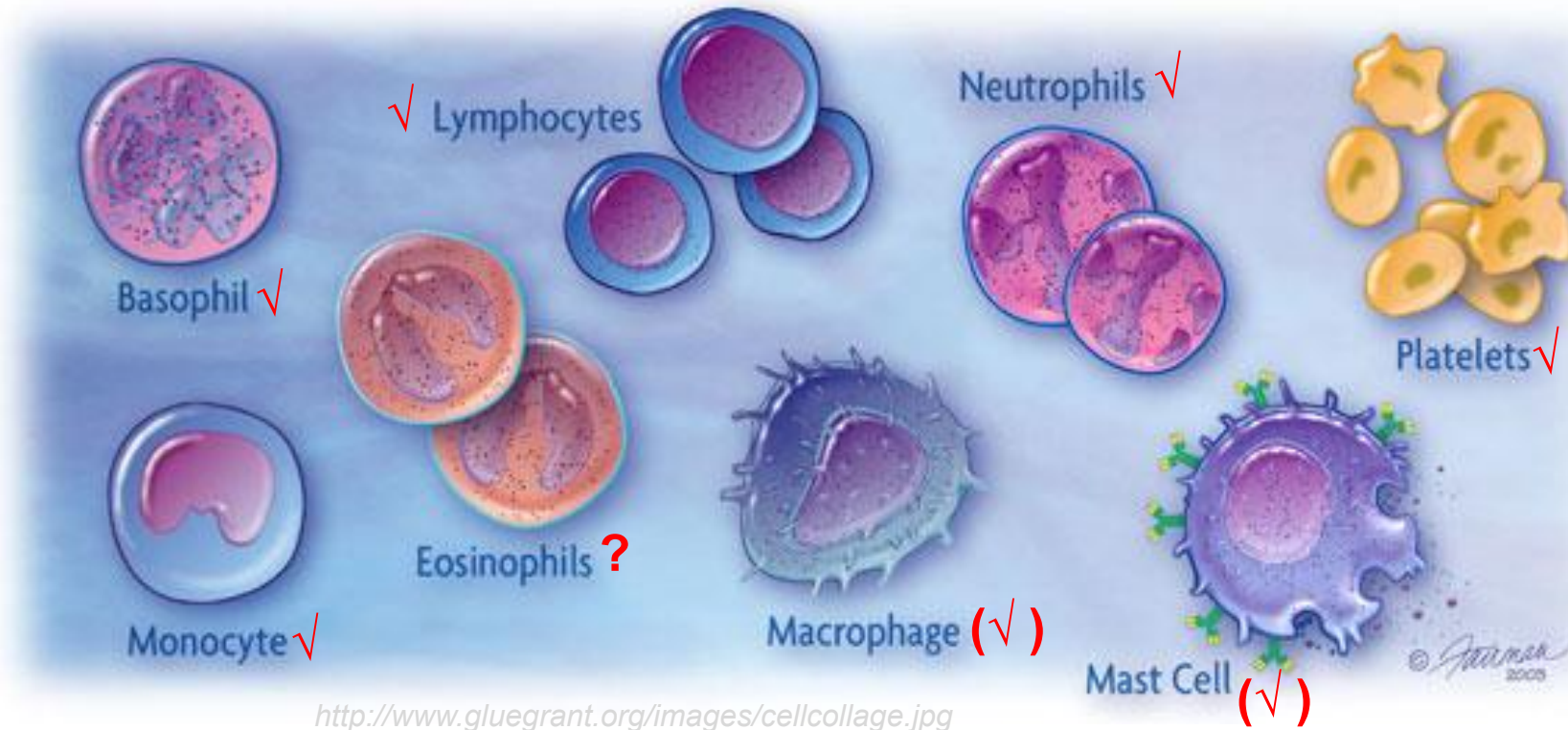


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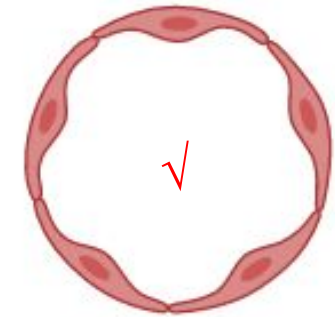
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- **JAK2V617F-mutant progenitors proliferate and further differentiate**
 - Progenitor differentiation may be skewed (towards myeloid, away from lymphoid differentiation)



Inflammatory cells in MF: are they part of the JAK2V617F clone?



<http://www.gluegrant.org/images/cellcollage.jpg>



Endothelial cells
(Blood vessel)

https://cdn.prod.website-files.com/621e95f9ac30687a56e4297e/64401622763e1e837b0490b9_vessel-cross-section-arteriole-endothelium.png

✓ = JAK2 V617F mutation **shown at DNA level**

- But JAK2V617F protein expression unclear (=> single-cell proteomics)
- Biologic contribution of each cell subtype unclear

BM stromal cells are negative for JAK2V617F

Patient no.	Diagnosis	<i>JAK2</i> ^{V617F} allele burden (%)	
		Granulocytes	MSCs
1	PMF	31	b.d. (<i>below detection limit</i>)
2	PMF	15	b.d.
3	PMF	23	b.d.
4	Post-PV MF	61	b.d.
5	Post-PV MF	98	b.d.
6	PV	100	b.d.
7	PV	66	b.d.

The JAK2 V617F vascular niche in MPN and its response to IFNa

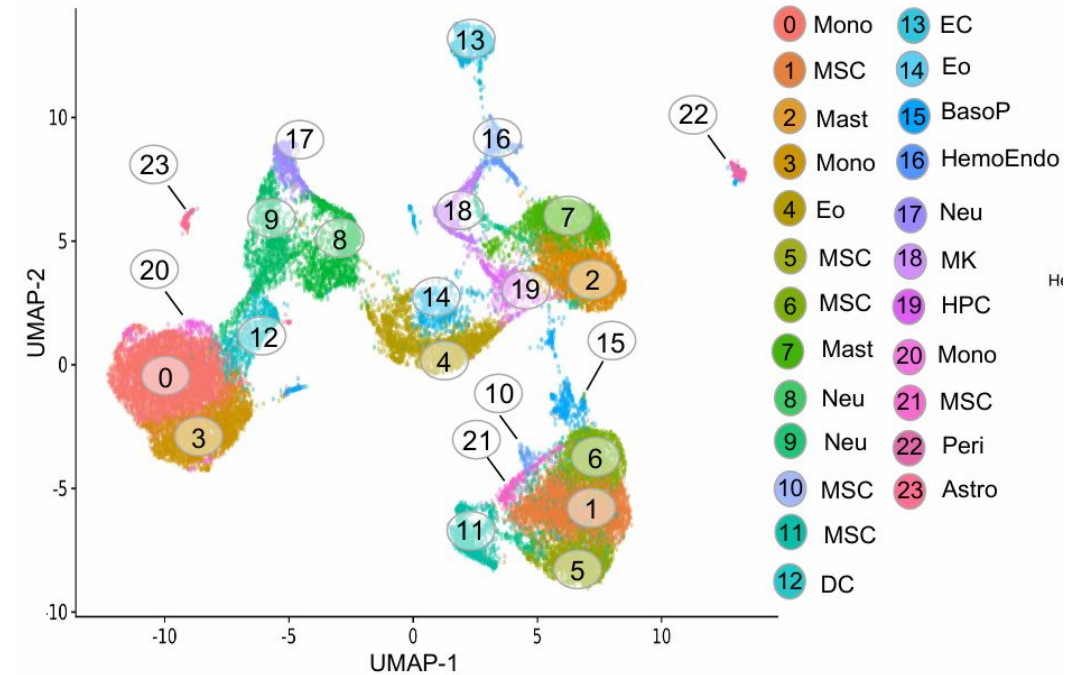
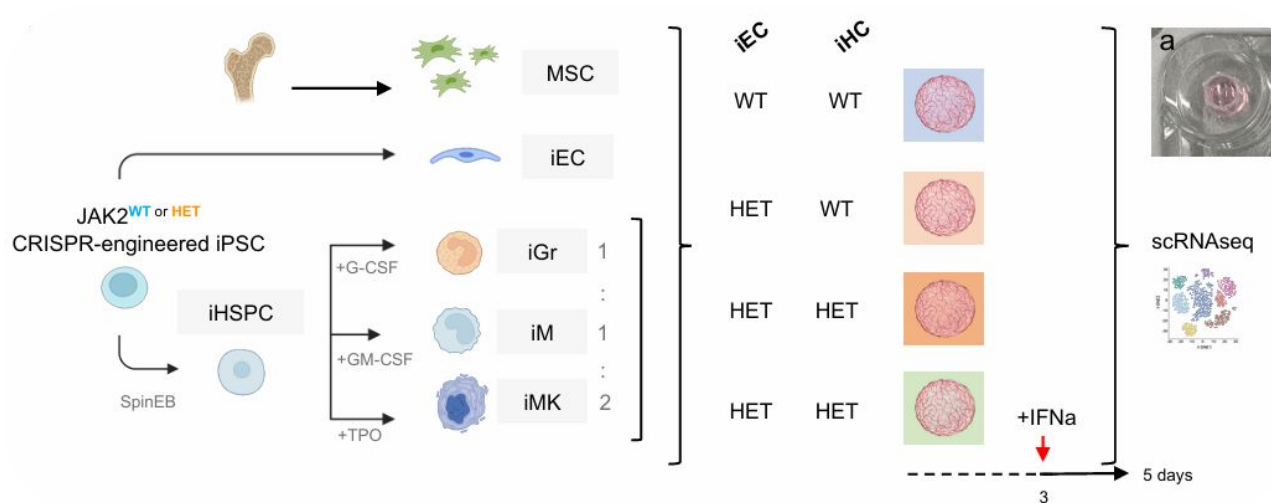


M. Caduc



M. de Toledo

- Recapitulating the MPN BM niche with iPSC-derived 3D cocultures (JAK2V617F in iHC +/- iEC)



The JAK2 V617F vascular niche in MPN and its response to IFN α

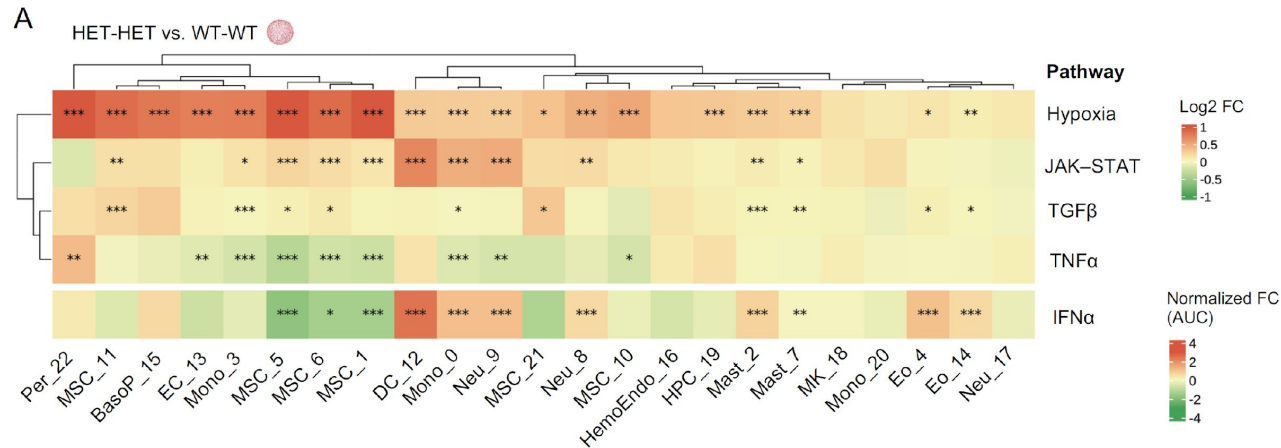


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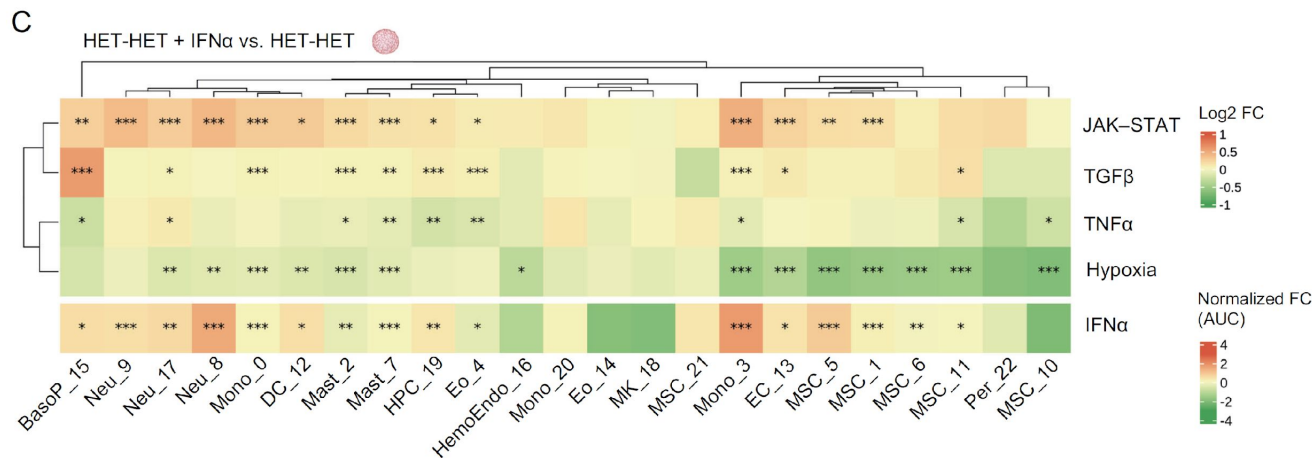


M. de Toledo

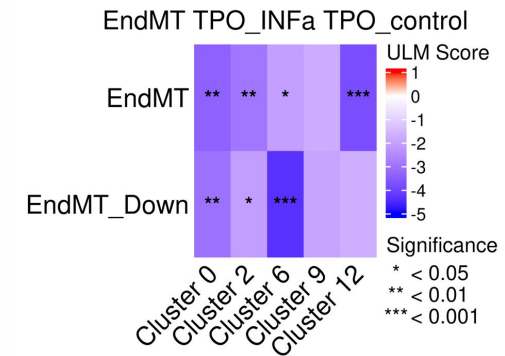
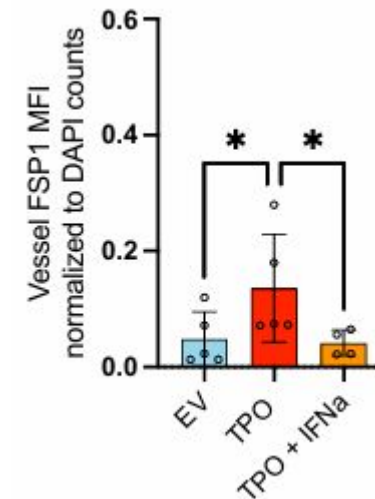
- Core MPN signatures such as hypoxia, JAK-STAT, and fibrosis (TGF β) recap. in 3D BM niches



- IFN α reverts the hypoxic and profibrotic phenotype *in vitro* and *in vivo*

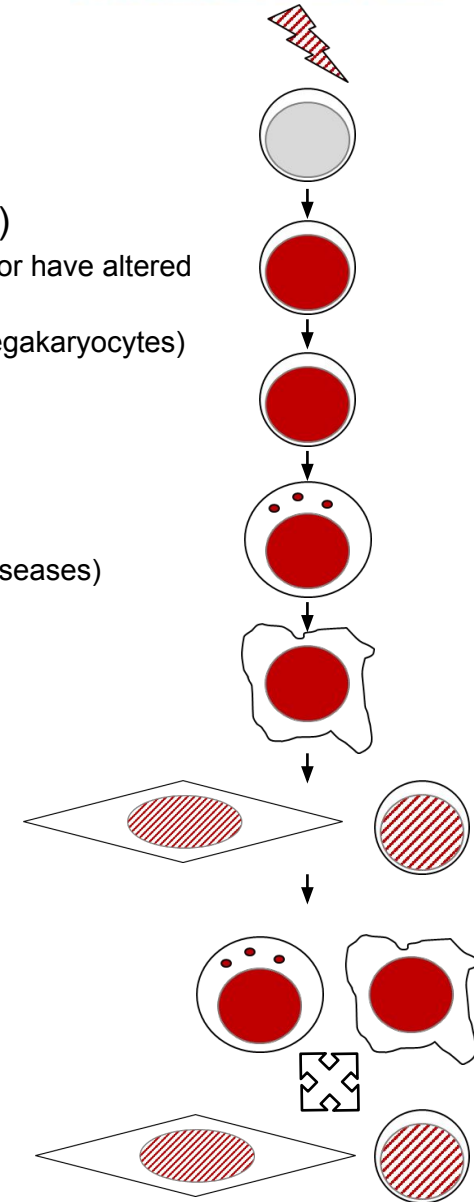


TPO mouse model

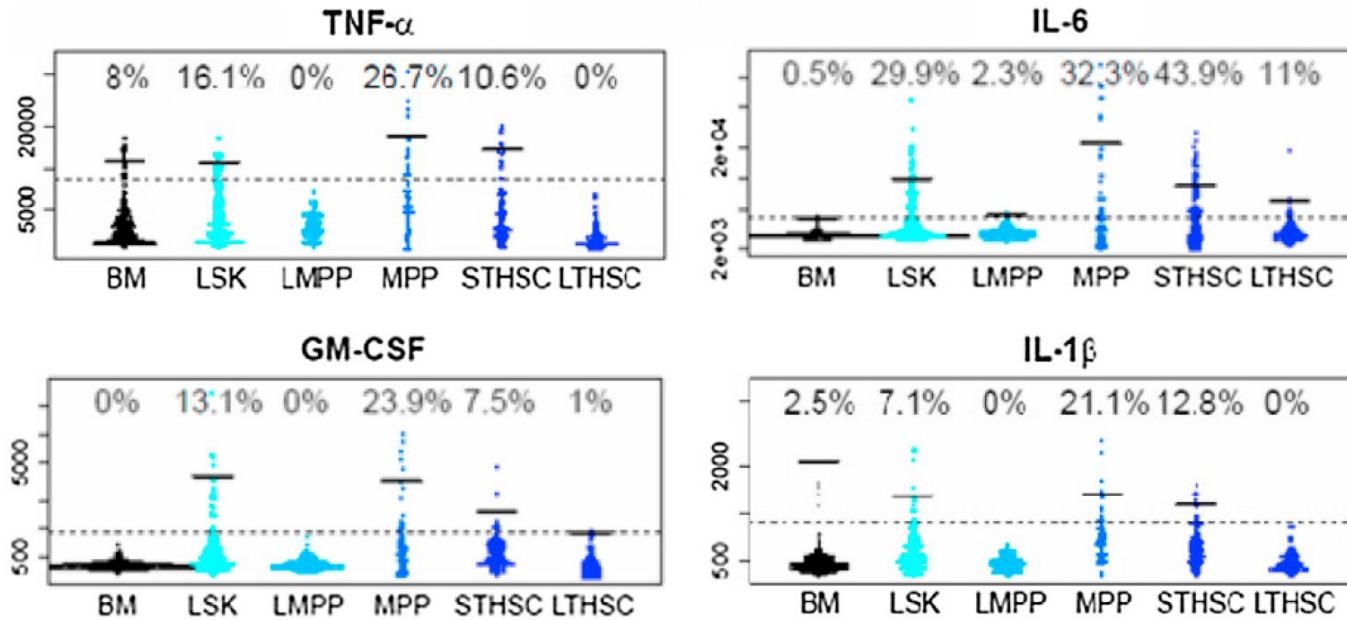


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- **JAK2V617F-mutant progenitors proliferate and further differentiate**
 - Progenitor differentiation may be skewed (towards myeloid, away from lymphoid differentiation)
- **JAK2V617F-mutant progeny produces local myeloid cytokines** (e.g. IL-1beta, IL-6, TNF)
 - JAK2V617F-mutant HSC and progenitors may enhance their cell division activity (e.g. proliferation, differentiation)
 - Clonal hematopoiesis (CHIP) for JAK2V617F may be detectable by PCR/NGS diagnostics (☐ Patient is at risk for MPN & cardiovascular diseases)

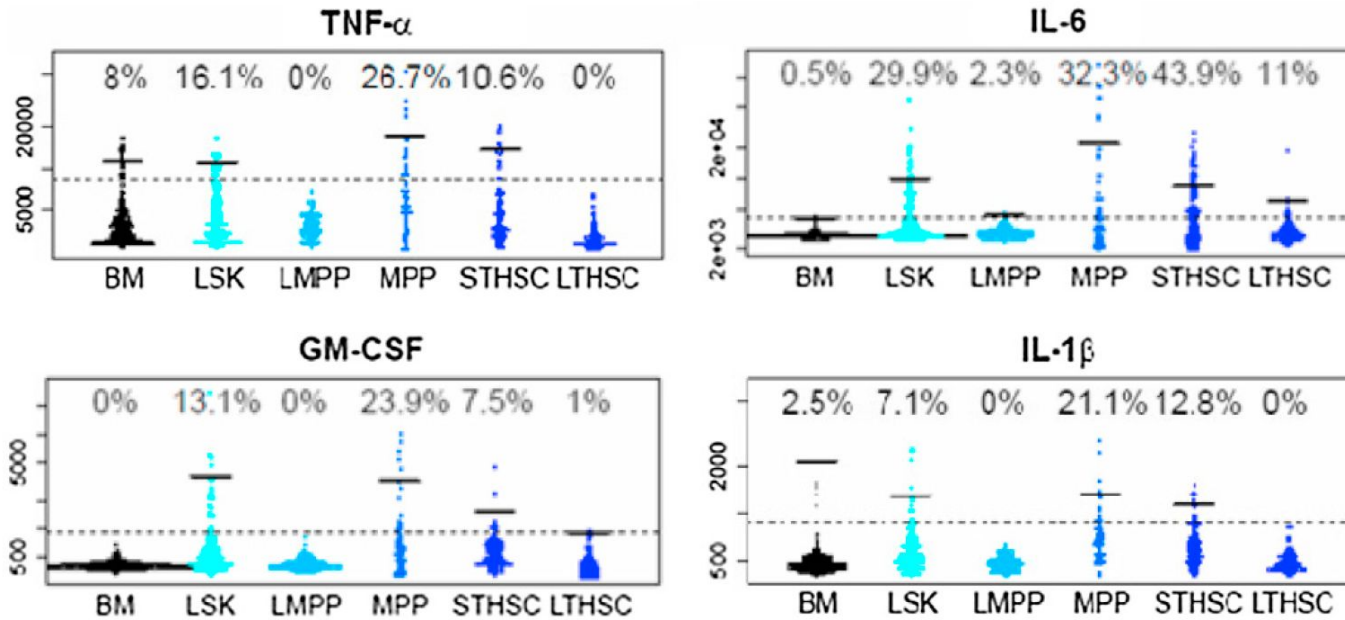


Single HSCs and MPPs produce inflammatory cytokines

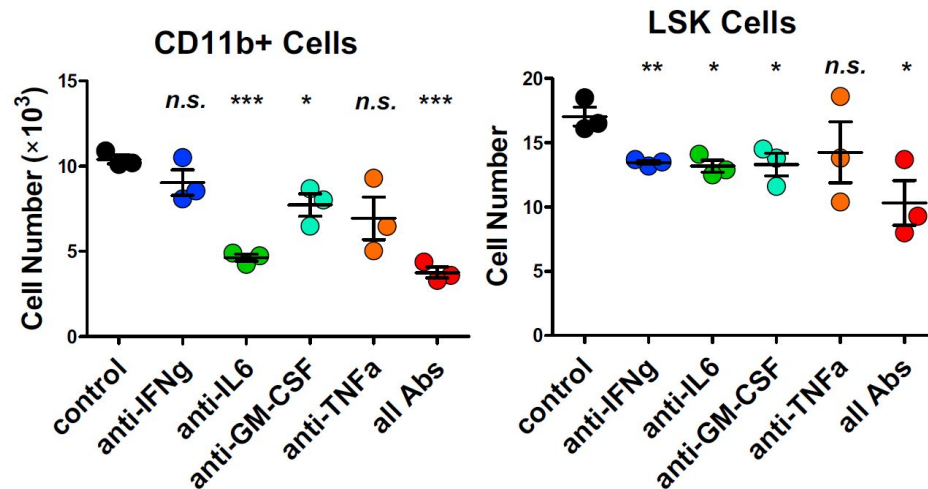


Single-cell proteomics

Single HSCs and MPPs produce inflammatory cytokines



Single-cell proteomics



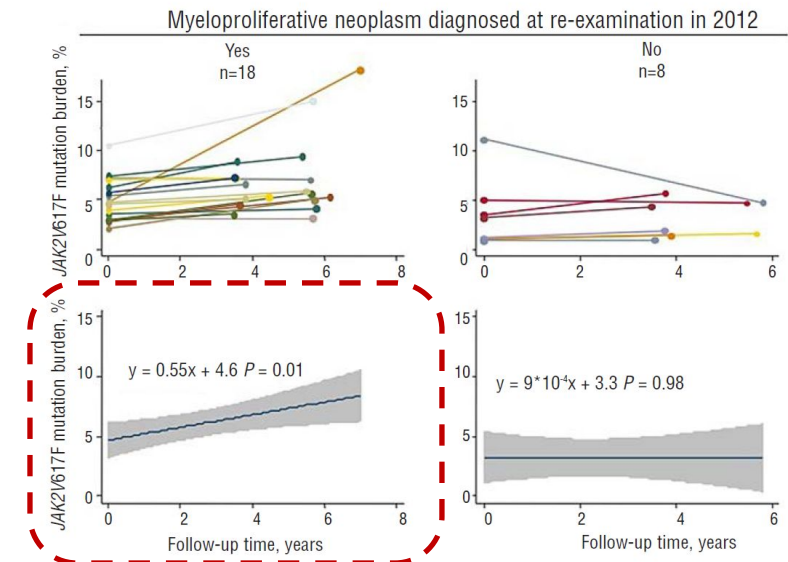
High risk of MPN in persons with clonal hematopoiesis (CHIP)

	JAK2 V617F (3.1%)	CALR (0.16%)	
		Type 1	Type 2
Number	613	24	8
Allele burden, %			
Mean (SE), range	2.1 (0.34), 0.010-96	6.2 (2.3), 0.020-44	11 (5.9), 0.013-38
<0.1, n (%)	255 (42)	5 (21)	1 (13)
0.1-0.99, n (%)	253 (41)	9 (38)	4 (50)
1-10, n (%)	75 (12)	4 (17)	0
>10, n (%)	30 (5)	6 (25)	3 (38)

Table III. Prevalent morbidity in the general population according to JAK2 V617F somatic mutation status.

Endpoints	JAK2 V617F somatic mutation status		Odds ratio (95% CI)
	Negatives (n = 49 420) cases/controls	Positives (n = 68) cases/controls	
Any cancer	6969/42 451	23/45	2.7 (1.6–4.6)
Haematological cancer	139/49 281	10/58	44 (22–90)
Myeloproliferative cancer	32/49 388	10/58	221 (100–487)
Ischaemic heart disease	2689/46 731	11/57	2.2 (1.1–4.4)
Myocardial infarction	1091/48 329	6/62	2.6 (1.1–6.3)
Venous thromboembolism	989/48 431	5/63	3.1 (1.3–7.9)
Pulmonary embolism	344/49 076	0/68	–
Deep venous thrombosis	729/48 691	5/63	4.6 (1.7–10.9)

Odds ratios were adjusted for sex, age, tobacco consumption, alcohol consumption, and body mass index at the time of blood sampling. CI = confidence interval.



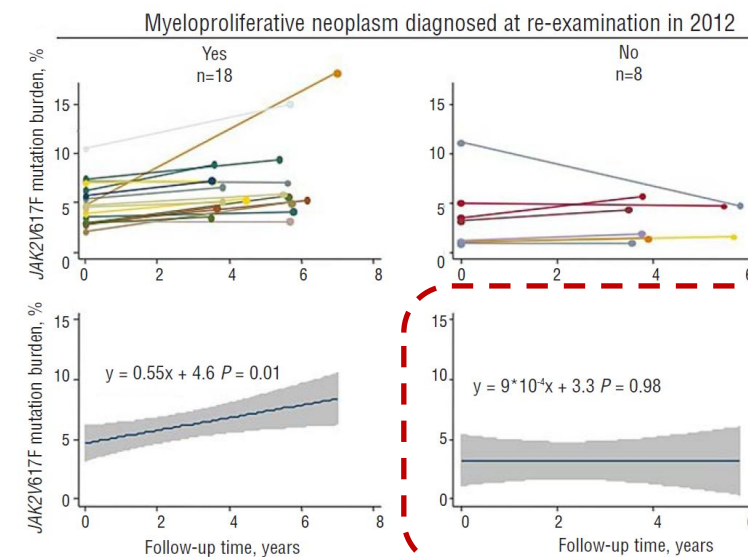
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Table III. Prevalent morbidity in the general population according to JAK2 V617F somatic mutation status.

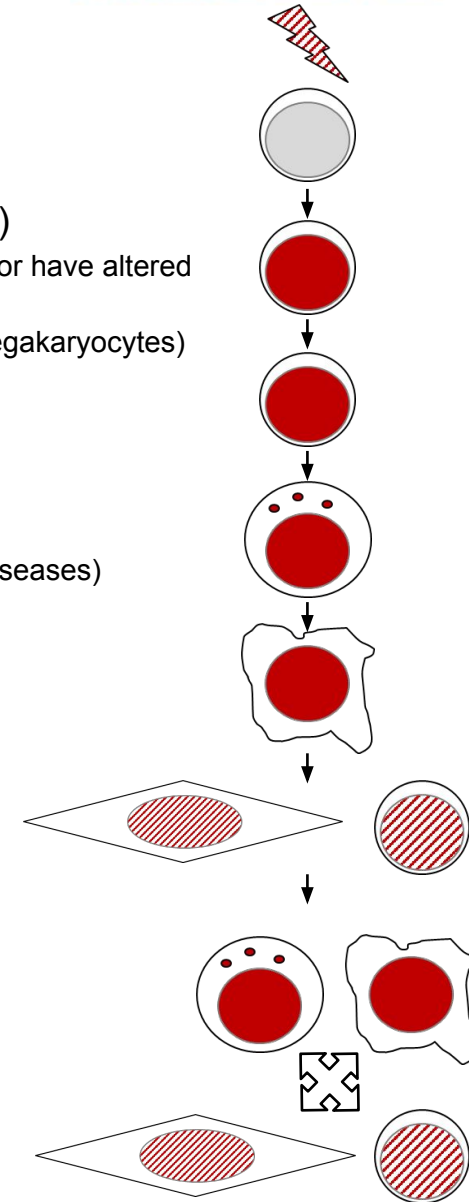
Endpoints	JAK2 V617F somatic mutation status		Odds ratio (95% CI)
	Negatives (n = 49 420) cases/controls	Positives (n = 68) cases/controls	
Any cancer	6969/42 451	23/45	2.7 (1.6–4.6)
Haematological cancer	139/49 281	10/58	44 (22–90)
Myeloproliferative cancer	32/49 388	10/58	221 (100–487)
Ischaemic heart disease	2689/46 731	11/57	2.2 (1.1–4.4)
Myocardial infarction	1091/48 329	6/62	2.6 (1.1–6.3)
Venous thromboembolism	989/48 431	5/63	3.1 (1.3–7.9)
Pulmonary embolism	344/49 076	0/68	–
Deep venous thrombosis	729/48 691	5/63	4.6 (1.7–10.9)

Odds ratios were adjusted for sex, age, tobacco consumption, alcohol consumption, and body mass index at the time of blood sampling. CI = confidence interval.



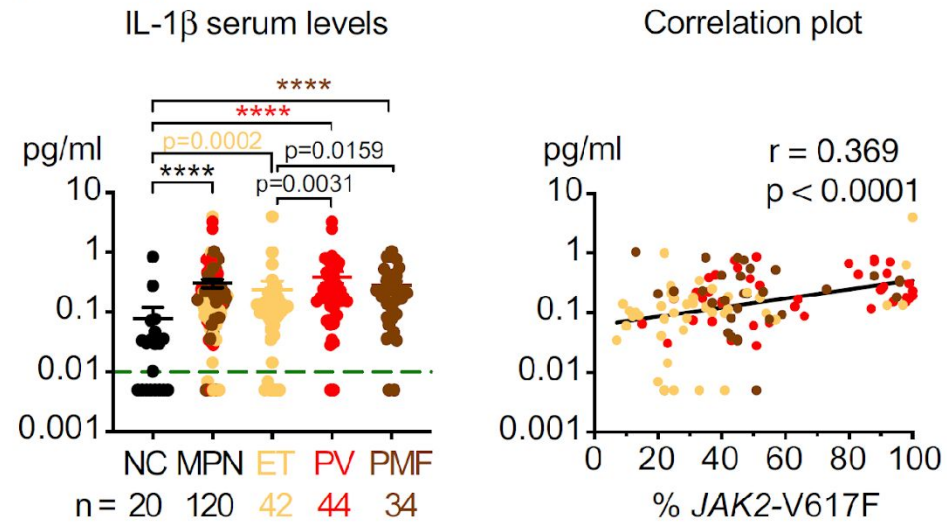
What is the evidence?

- **HSC** in the bone marrow (BM) **acquires a somatic JAK2V617F** (or CALR or MPL) driver mutation
 - This has been described to occur very early in life (e.g. in utero or early childhood)
 - JAK2V617F has been shown to induce cycling of HSCs
- **JAK2V617F-mutant HSC divide** symmetrically or asymmetrically (distribution and influential factors are still unclear)
 - Symmetrical: JAK2V617F-mutant HSC generates another JAK2V617F pos HSC (unclear whether these 2° HSCs are biologically identical or have altered function)
 - Asymmetrical: JAK2V617F-mutant HSC differentiates into MPPs/CMPs/MEPs/GMPs (HSC differentiation may be skewed, e.g. towards megakaryocytes)
- **JAK2V617F-mutant progenitors proliferate and further differentiate**
 - Progenitor differentiation may be skewed (towards myeloid, away from lymphoid differentiation)
- JAK2V617F-mutant progeny produces **local myeloid cytokines** (e.g. IL-1beta, IL-6, TNF)
 - JAK2V617F-mutant HSC and progenitors may enhance their cell division activity (e.g. proliferation, differentiation)
 - Clonal hematopoiesis (CHIP) for JAK2V617F may be detectable by PCR/NGS diagnostics (☐ Patient is at risk for MPN & cardiovascular diseases)
- JAK2V617F-mutant **differentiated cells accumulate in the peripheral blood** and cause **full-blown MPN**
 - Systemic cytokines may cause chronic inflammatory symptoms (fever, night sweats, weight loss, fatigue, pruritus, ...)
 - Thrombocytosis, erythrocytosis, leukocytosis (mostly neutrophilia, monocytosis), splenomegaly
 - Highly increased risk of cardiovascular complications (e.g. thrombosis, bleeding, organ damage)

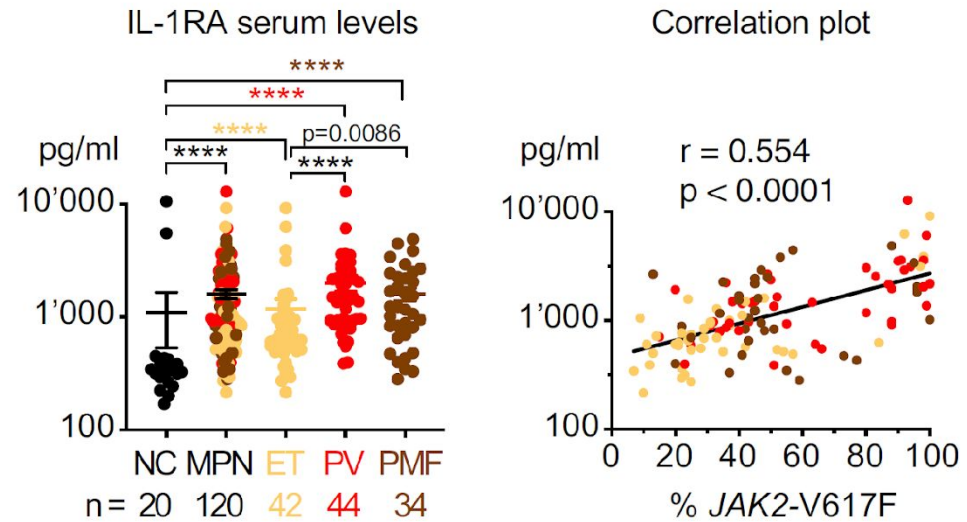


IL-1beta is required for JAK2V617F-mediated MPN in mice

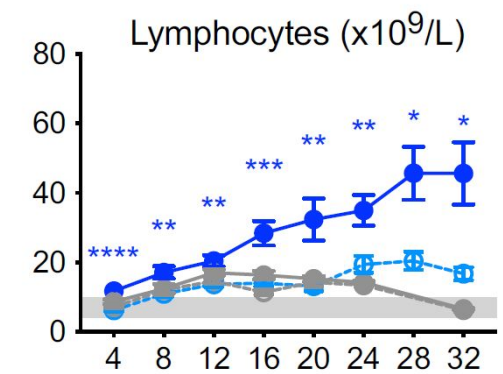
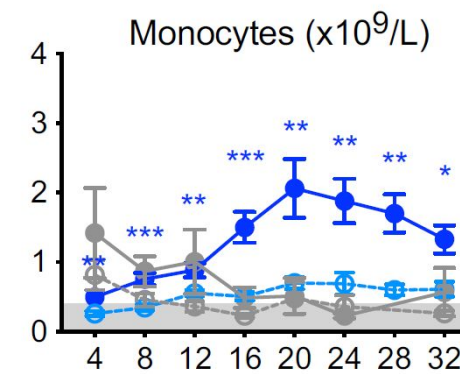
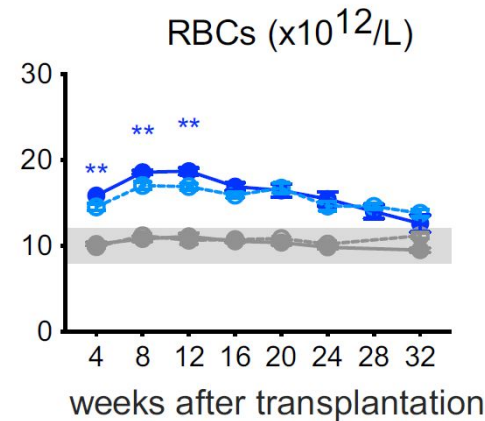
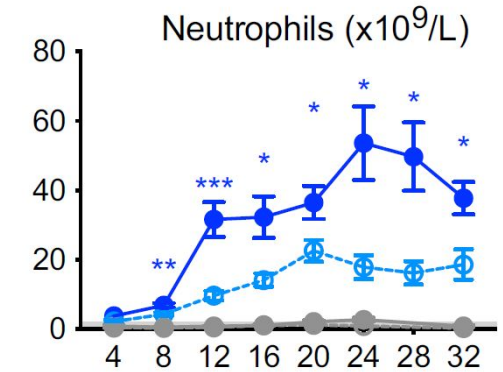
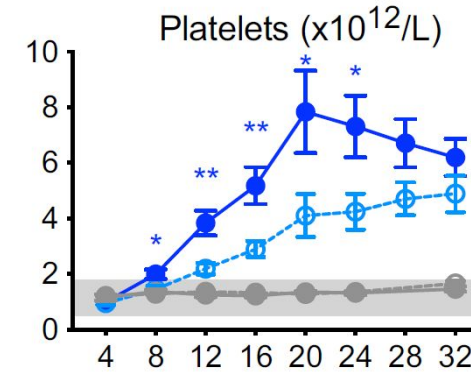
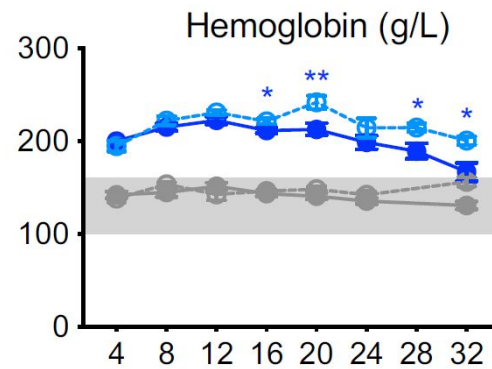
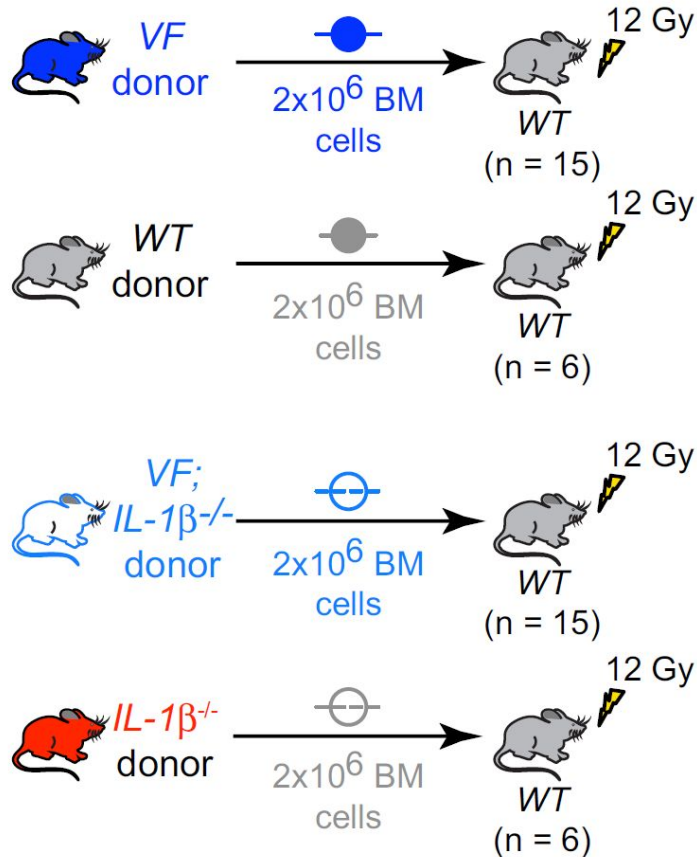
a IL-1 β and JAK2-V617F allele burden



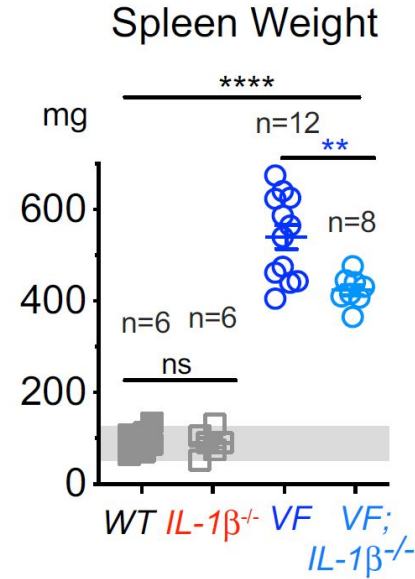
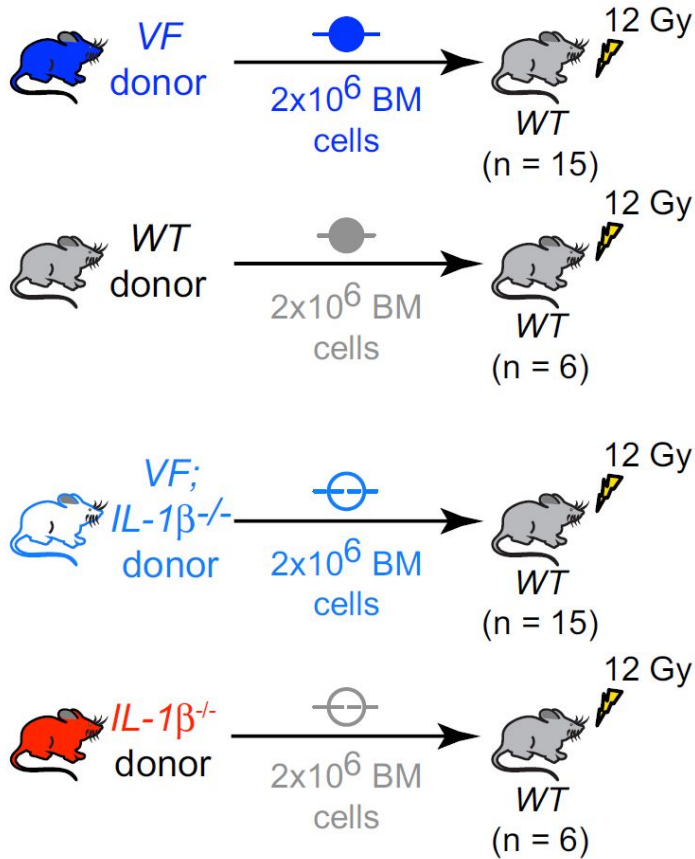
b IL-1RA and JAK2-V617F allele burden



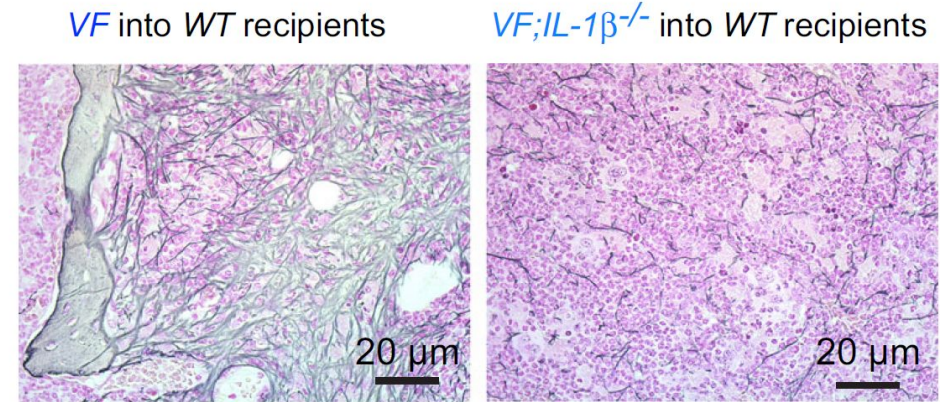
IL-1beta is required for JAK2V617F-mediated MPN in mice



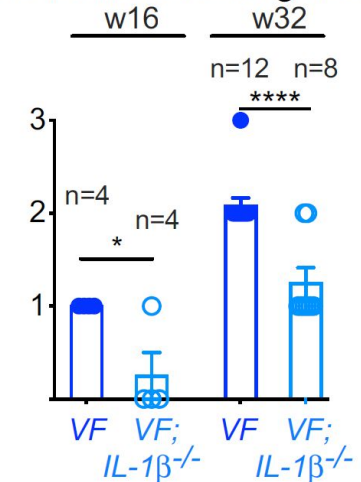
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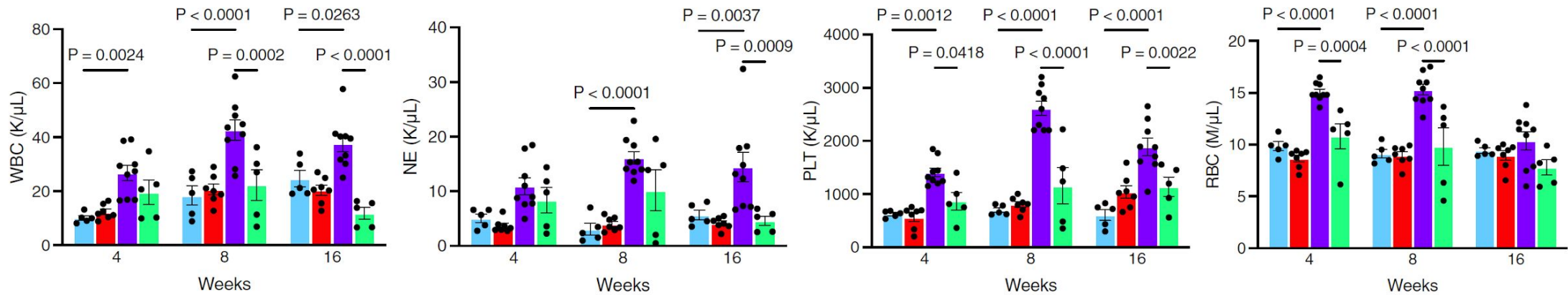
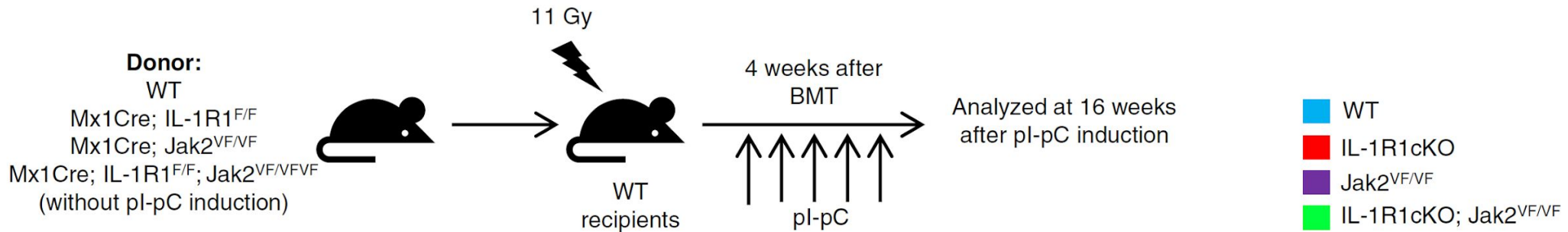
Bone marrow fibrosis at 32 weeks post Tx



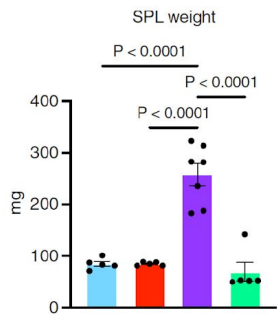
Reticulin fibrosis grade



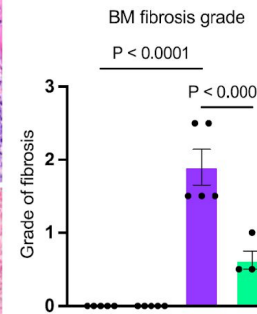
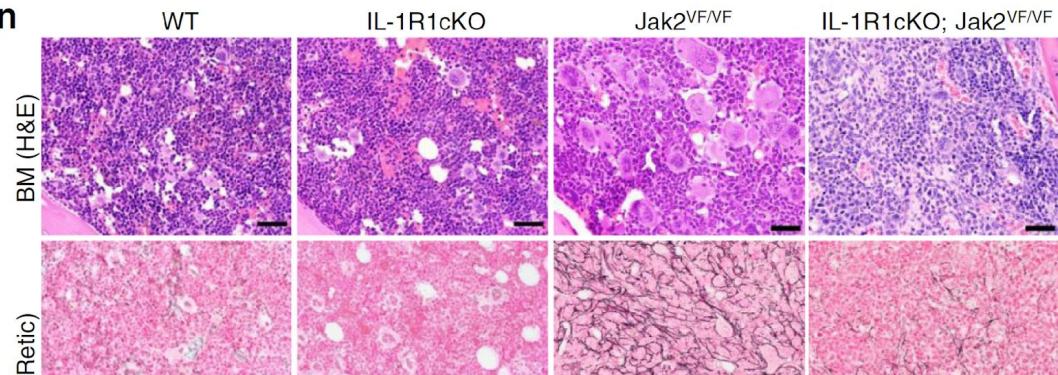
IL-1R1 is essential for JAK2V617F-mediated MPN in mice



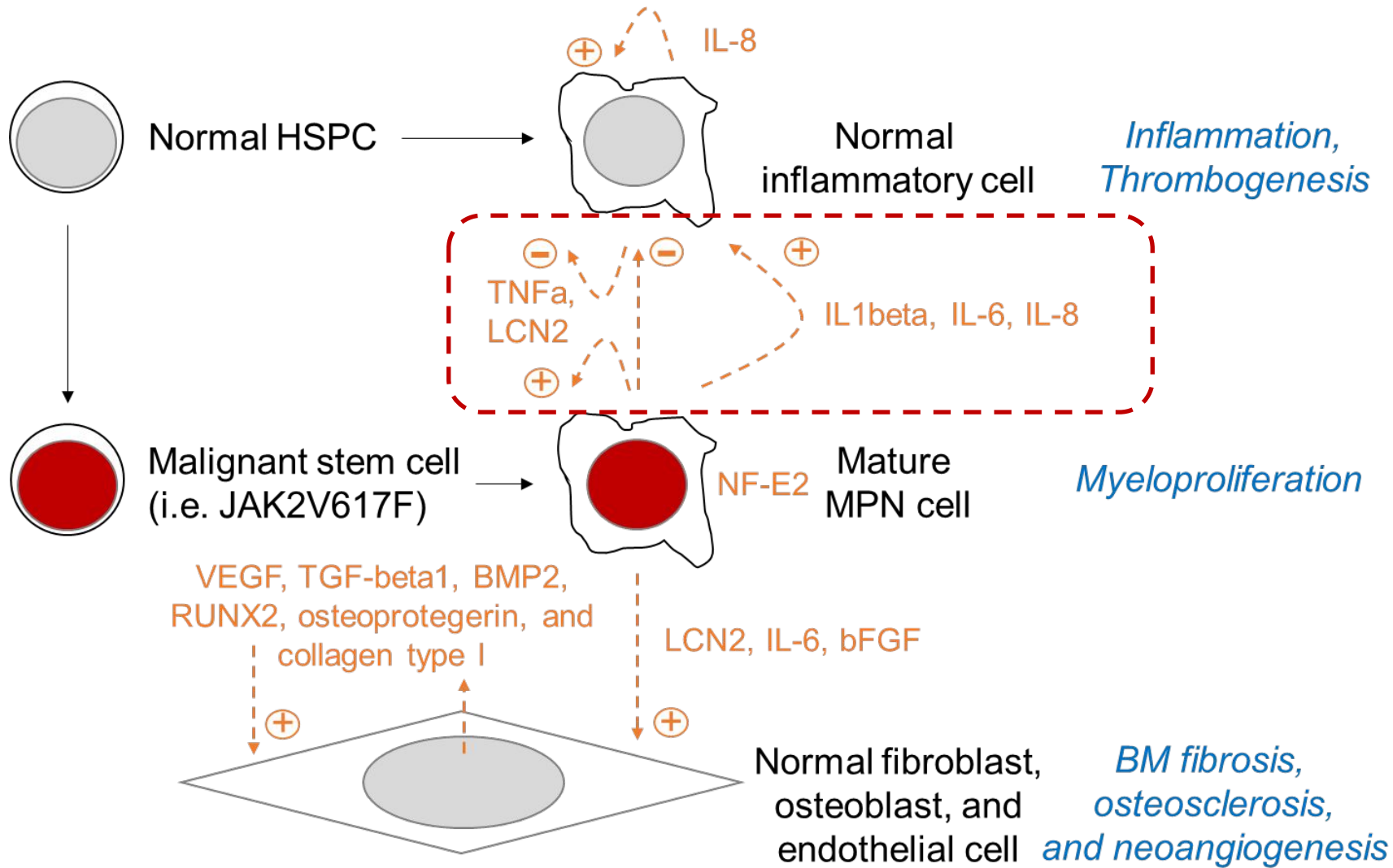
m



n



Inflammation confers selective advantage to the malignant clone



Inflammatory cytokine serum levels are increased in MF

Table 2. Cytokines Whose Plasma Levels Are Abnormally Increased (or decreased) in PMF and Their Relationship With Age, Sex, and Clinically Relevant Disease Features at Time of First Referral at the Mayo Clinic

Cytokines (pg/mL)	Controls (n = 35)		All Patients With PMF (N = 127)		P	P									
	Median	Range	Median	Range		Constitutional Symptoms	RBC Transfusion Dependency	Spleen > 10 cm	WBC > 10 × 10 ⁹ /L	Platelets > 450 × 10 ⁹ /L	Platelets < 100 × 10 ⁹ /L	JAK2V617F Positivity	Age	Sex	
IL-1β	4	0-49	10.8	0-3,576	.02	.81	.68	.21	.87	.64	.3	.35	.12	.05	
IL-1RA	203	2-419	552	37-9,991	< .001	.84	.62	.07	.68	.27	.47	< .001	.14	.26	
IL-2R	217	0-507	556	91-3,956	< .001	.24	.001	.17	.006	.76	.68	.008	.06	.01†	
IL-6	0.6	0-9	6.3	0-186	< .001	.03	.05	.09	.34	.13	.35	.007	.05	.12	
IL-8	3.3	0-18	14.3	0-1,156	< .001	.004	.03	.35	.07	.58	.26	.63	.1	.04†	
IL-10	4.8	2.3-51	12.5	2-2,009	< .001	.48	.02	.34	.73	.23	.31	.61	.55	.22	
IL-12	100	35-182	192	18-1,883	< .001	.55	.14	.28	.80	.99	.04*	.02	.34	.43	
IL-13	0	0-0	0	0-4,909	.001	.9	.06	.46	.46	.87	.97	.48	.09	.11	
IL-15	0	0-38	0	0-2,671	.03	.8	.61	.46	.86	.95	.90	.85	.16	.02†	
TNF-α	0	0-15	0	0-400	.02	.97	.13	.09	.60	.48	.95	.58	.97	.08	
G-CSF	33	0-373	45	0-888	.007	.1	.91	.99	.72	.23	.08	.66	.34	.38	
IFN-α	27.6	0-96	42	0-1,021	.02	.4	.11	.96	.33	.59	.95	.83	.99	.06	
IFN-γ	5.5	0-23	0	0-683	.02*	.55	.8	.55	.5	.21	.53	.12	.91	.52	
MIP-1α	0	0-112	25.4	0-1,305	< .001	.55	.03	.05	.21	.87	.74	.98	.14	.001†	
MIP-1β	21.8	4.4-91	65.7	0-1,935	< .001	.67	.3	.16	.69	.12	.03	.05	.07	.74	
HGF	129	0-433	391	0-11,572	< .001	.14	.9	.02	.02	.27	.39	.003	.54	.76	
IP-10	22	4-97	72	5.3-755	< .001	.08	.2	.19	.02	.11	.006	< .001	< .001	.73	
MIG	19.4	0-86	49	0-971	< .001	.48	.3	.08	.76	.44	.28	.01	.12	.04†	
MCP-1	173	61-342	222	62-1,705	.001	.19	.009	.77	.87	.20	.07	.27	.68	.24	
VEGF	1	0-2.7	2.3	0-47	< .001	.42	.42	.23	.72	.87	.27	.89	.31	.32	

Inflammatory cytokine serum levels are increased in MF

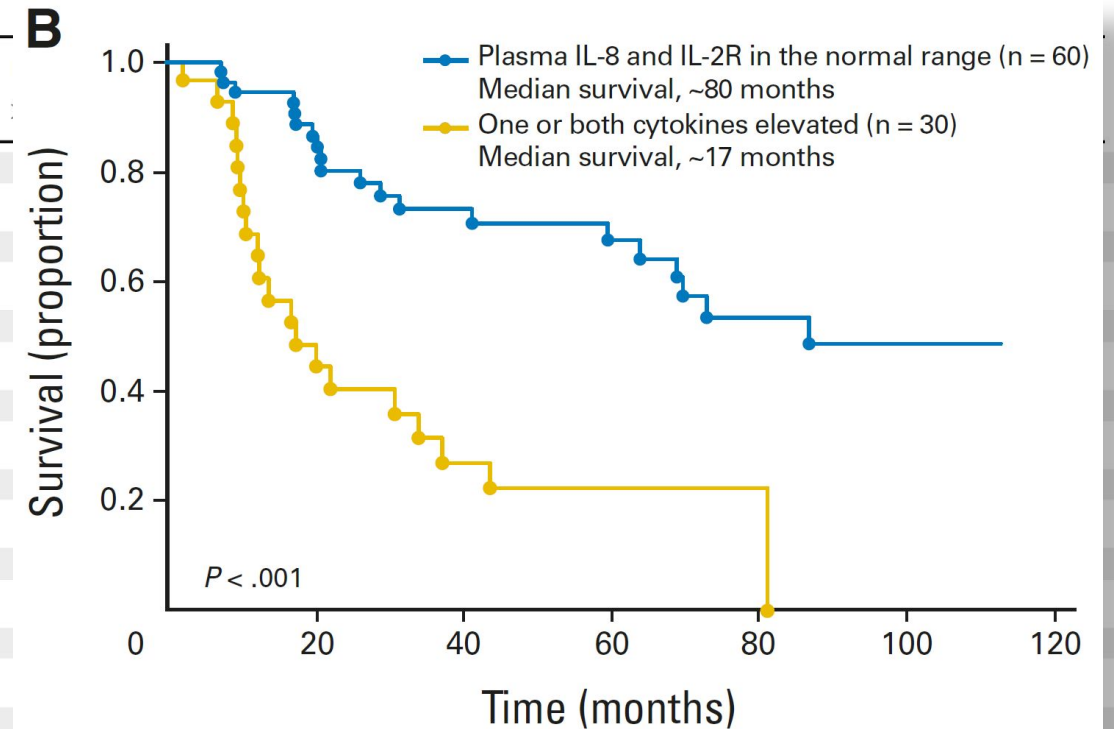
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IL-13	0	0-0	0	0-4,909	.001	.9	.06	.46	.46	.87	.97	.48	.09	.11
IL-15	0	0-38	0	0-2,671	.03	.8	.61	.46	.86	.95	.90	.85	.16	.02†
TNF-α	0	0-15	0	0-400	.02	.97	.13	.09	.60	.48	.95	.58	.97	.08
G-CSF	33	0-373	45	0-888	.007	.1	.91	.99	.72	.23	.08	.66	.34	.38
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Inflammatory cytokine serum levels are increased in MF

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IL-8	3.3	0-18	14.3	0-1,156	< .001	.004	.03
IL-10	4.8	2.3-51	12.5	2-2,009	< .001	.48	.02
IL-12	100	35-182	192	18-1,883	< .001	.55	.14
IL-13	0	0-0	0	0-4,909	.001	.9	.06
IL-15	0	0-38	0	0-2,671	.03	.8	.61
TNF- α	0	0-15	0	0-400	.02	.97	.13
G-CSF	33	0-373	45	0-888	.007	.1	.91
IFN- α	27.6	0-96	42	0-1,021	.02	.4	.11
IFN- γ	5.5	0-23	0	0-683	.02*	.55	.8
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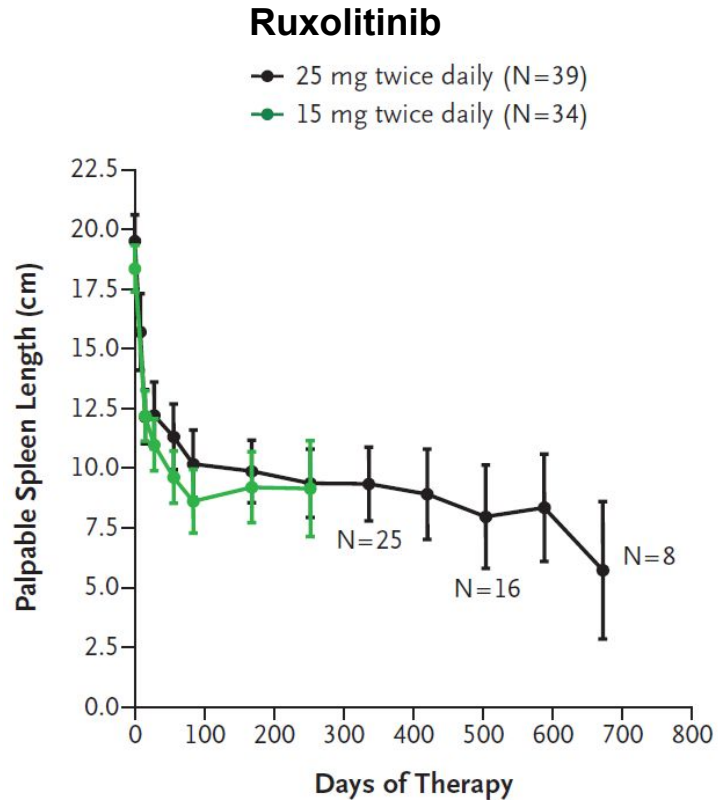
.02	.02	.27	.39	.003	.54	.76
.19	.02	.11	.006	< .001	< .001	.73
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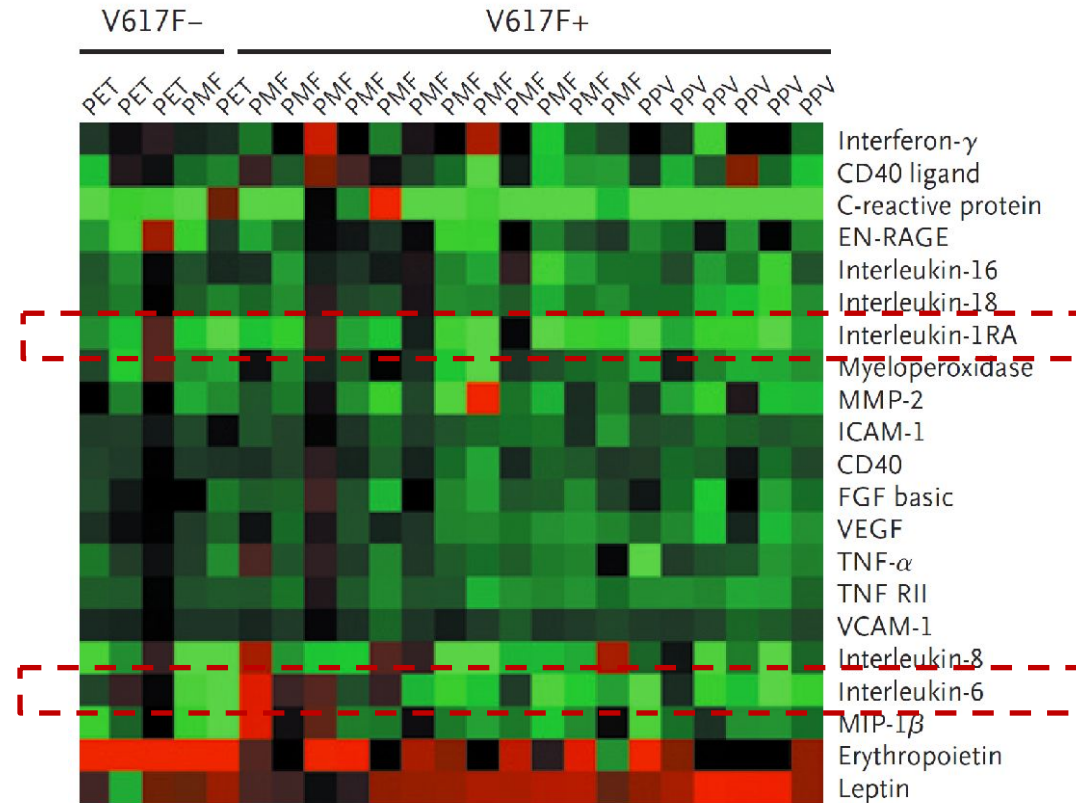
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JAK inhibitors may ameliorate MPN (in part via IL-6 decrease?)



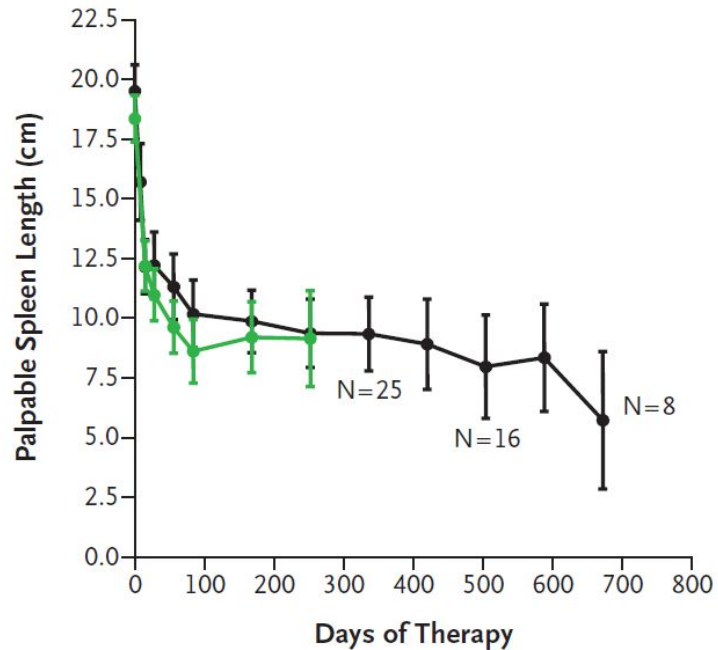
Patients with Myelofibrosis, Day 28 vs. Baseline



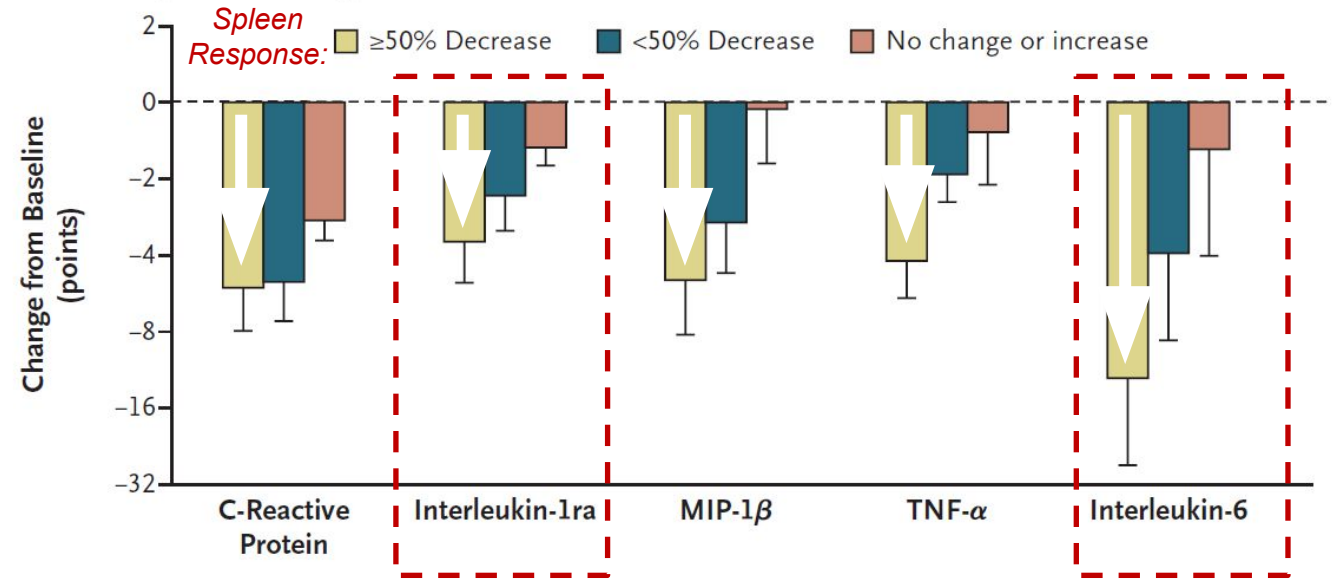
JAK inhibitors may ameliorate MPN (in part via IL-6 decrease?)

Ruxolitinib

- 25 mg twice daily (N=39)
- 15 mg twice daily (N=34)

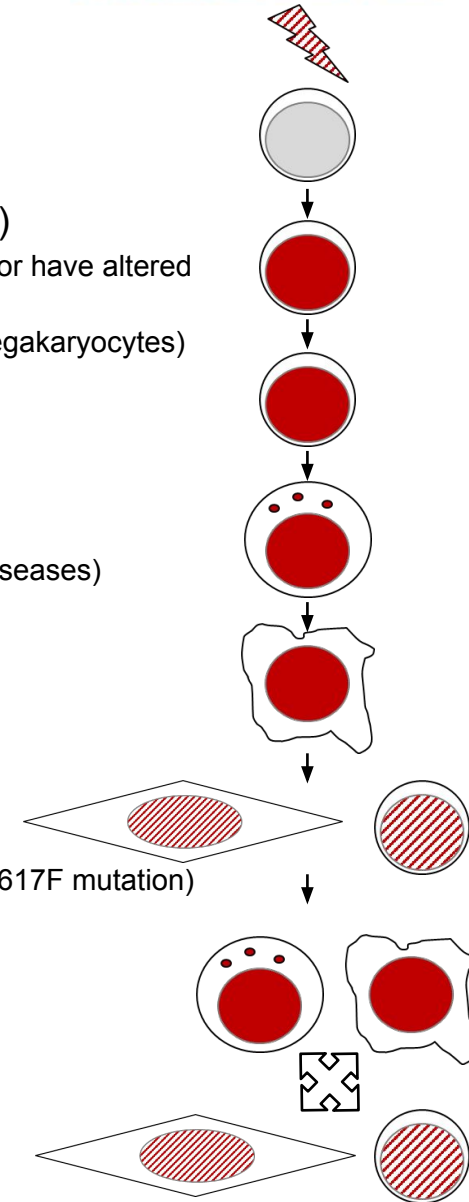


Change in Cytokine Level, 6 Cycles of Therapy

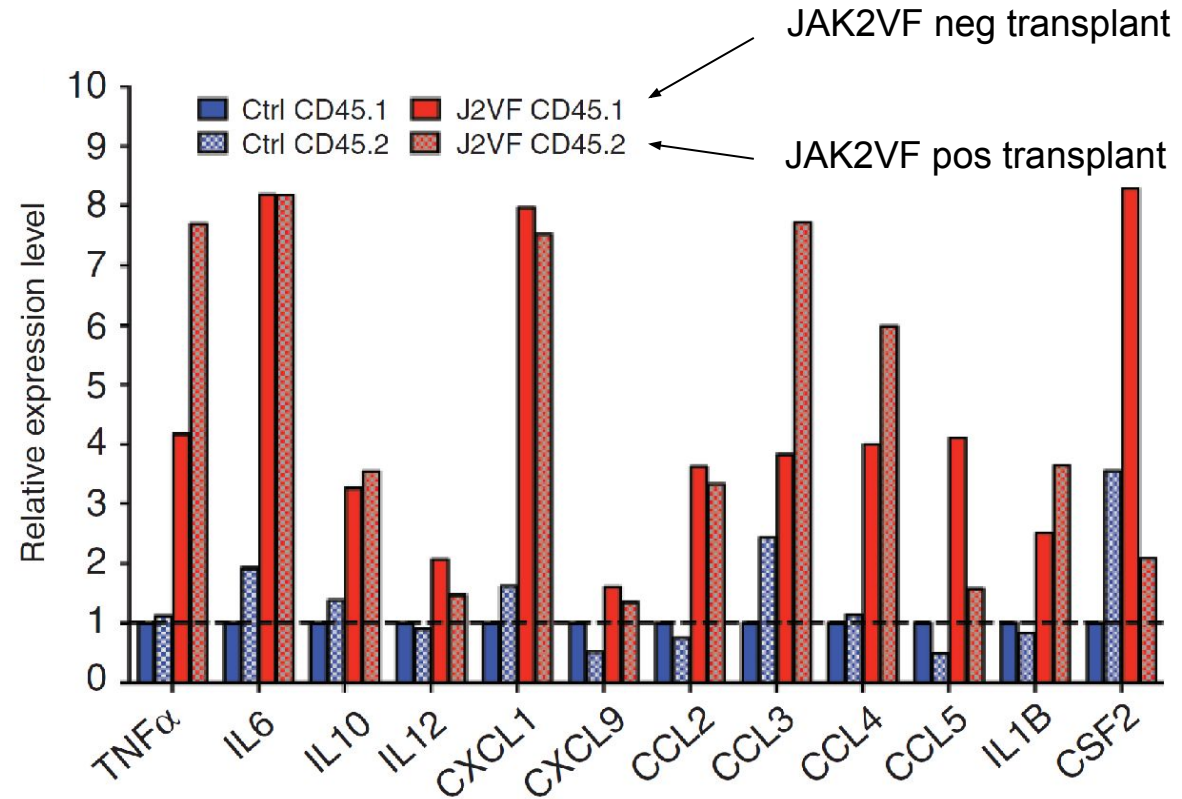
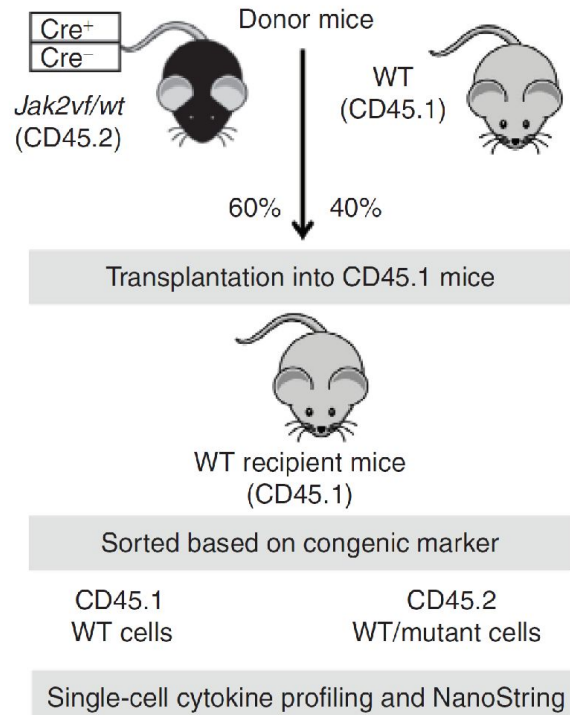


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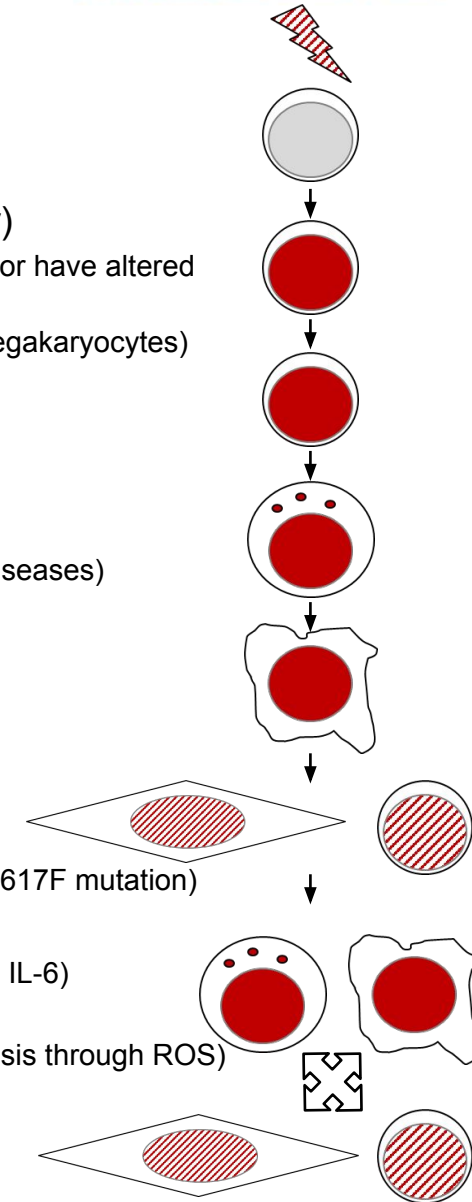


JAK2V617F clone alters normal hematopoiesis



What is the evidence?

- **HSC** in the bone marrow (BM) **acquires a somatic JAK2V617F** (or CALR or MPL) driver mutation
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 - These processes induce selection against TP53 WT in the hematopoietic compartment, favoring emergence of TP53 mutations (mutagenesis through ROS)
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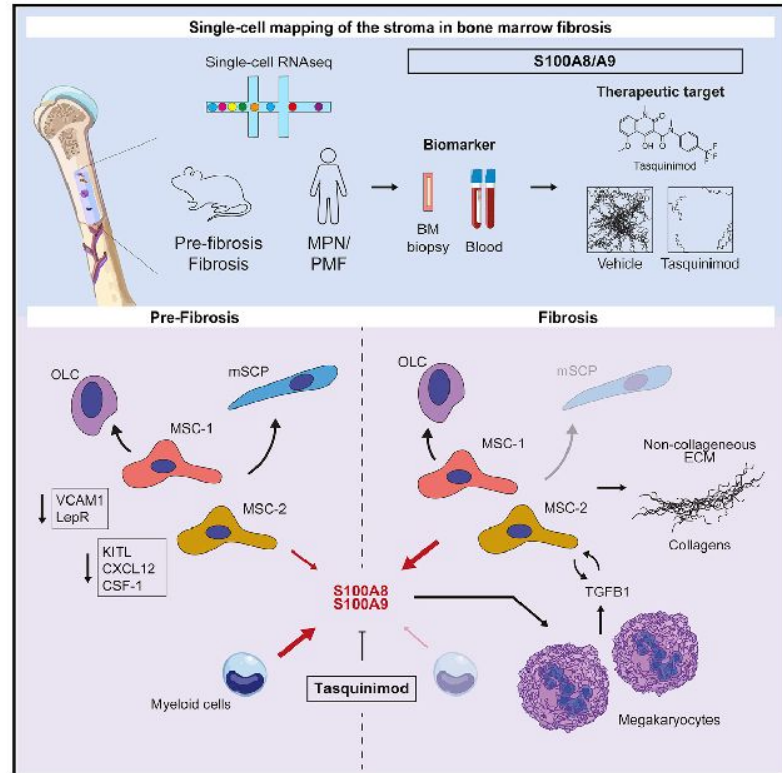


Article

Cell Stem Cell

Heterogeneous bone-marrow stromal progenitors drive myelofibrosis via a druggable alarmin axis

Graphical Abstract



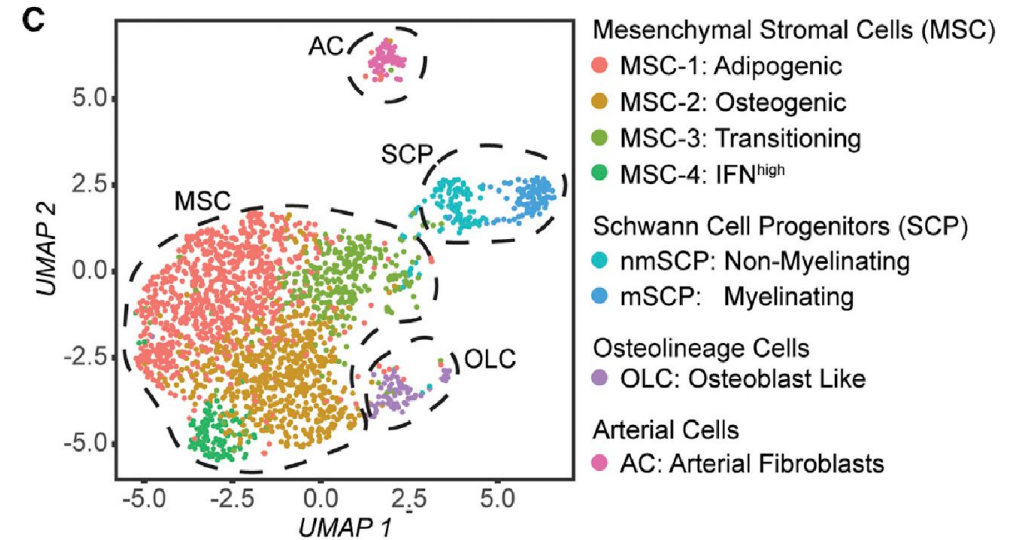
Nils B. Leimkühler,^{1,14} Hélène F.E. Gleitz,^{1,14} Li Ronghui,^{2,14} Inge A.M. Snoeren,¹ Stijn N.R. Fuchs,¹ James S. Nagai,² Bella Banjanin,¹ King H. Lam,³ Thomas Vogl,⁴ Christoph Kuppe,⁵ Ursula S.A. Stalman,¹ Guntram Büsche,⁶ Hans Kreipe,⁶ Ines Gütgemann,⁷ Philippe Krebs,⁸ Yara Banz,⁸ Peter Boor,⁹ Evelyn Wing-Ying Tai,⁹ Tim H. Brümmendorf,¹⁰ Steffen Koschmieder,¹⁰ Martina Crysandt,¹⁰ Eric Bindels,¹ Rafael Kramann,^{5,13,15} Ivan G. Costa,^{2,15} and Rebekka K. Schneider^{1,11,12,15,16,*}

Correspondence

reschneider@ukaachen.de

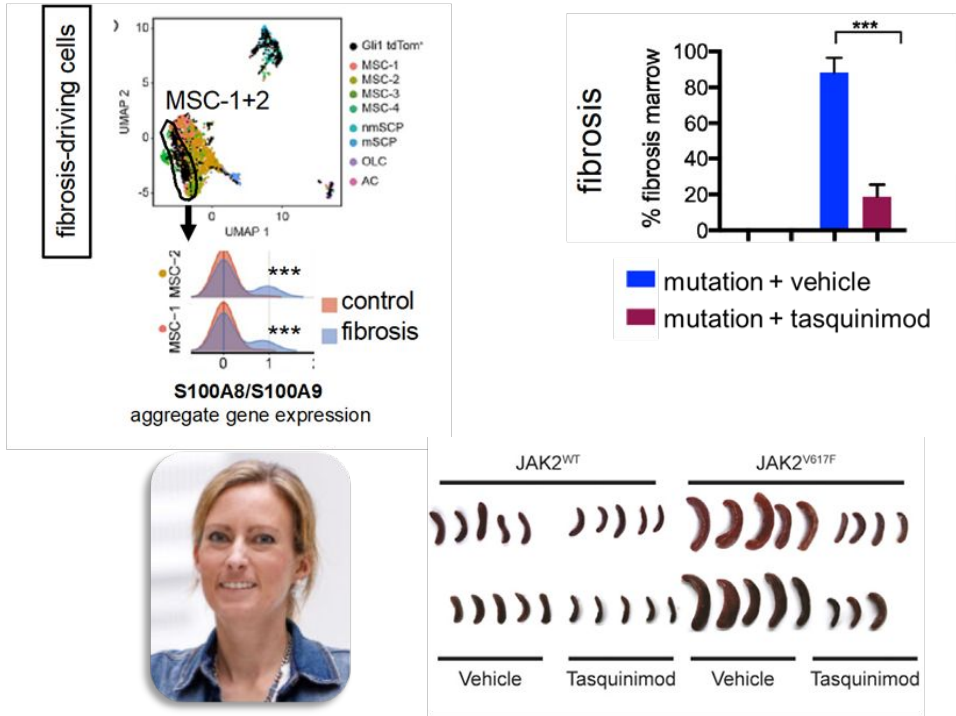
In Brief

Leimkühler and colleagues demonstrate that mesenchymal stromal progenitor cells are fibrosis-driving cells in mice and patients, that inflammation in the bone-marrow stroma precedes TGF-β signaling-driven fibrosis, and that the alarmin heterocomplex S100A8/S100A9 holds promise as MPN progression marker and therapeutic target.



Early interventions: Role of alarmins S100A8/A9 in MPN initiation

Target Identification (S100A8/A9) and Target Validation (Tasquinimod)¹



Rebekka Schneider
Aachen

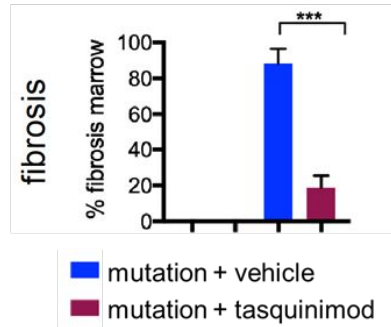
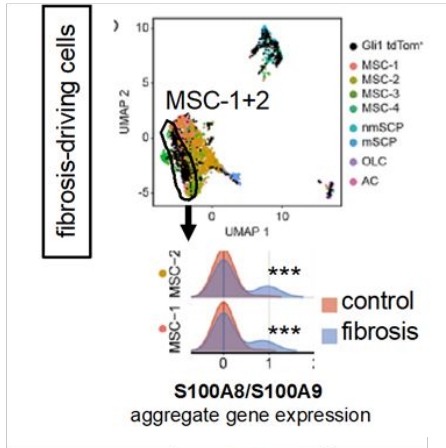
Tasquinimod is disease-modifying

- anti-fibrotic,
- anti-cytopenic,
- splenomegaly-reducing

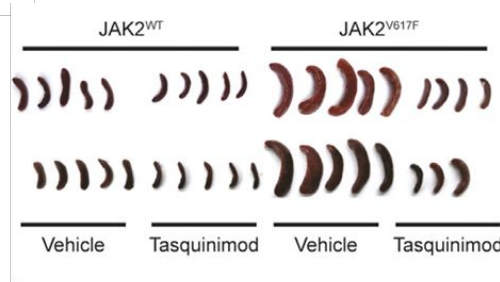
¹Leimkühler, Gleitz, Ronghui et al Cell Stem Cell 2021

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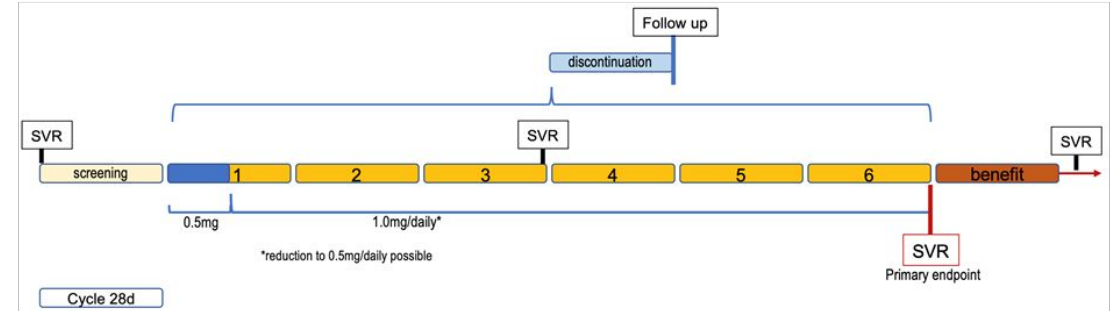


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Proof-of-concept Phase II Clinical Trial – TasquForce MPN

Patient cohort: heavily pretreated patients with MPN and bone marrow fibrosis



Peter te Boekhorst
Rotterdam



Martina Crysandt
Aachen

Funding:



Sponsor:



□ Clinical has recently started...

¹Leimkühler, Gleitz, Ronghui et al Cell Stem Cell 2021

Summary and Outlook

Aspects of Inflammation in MPN

Pathogenesis

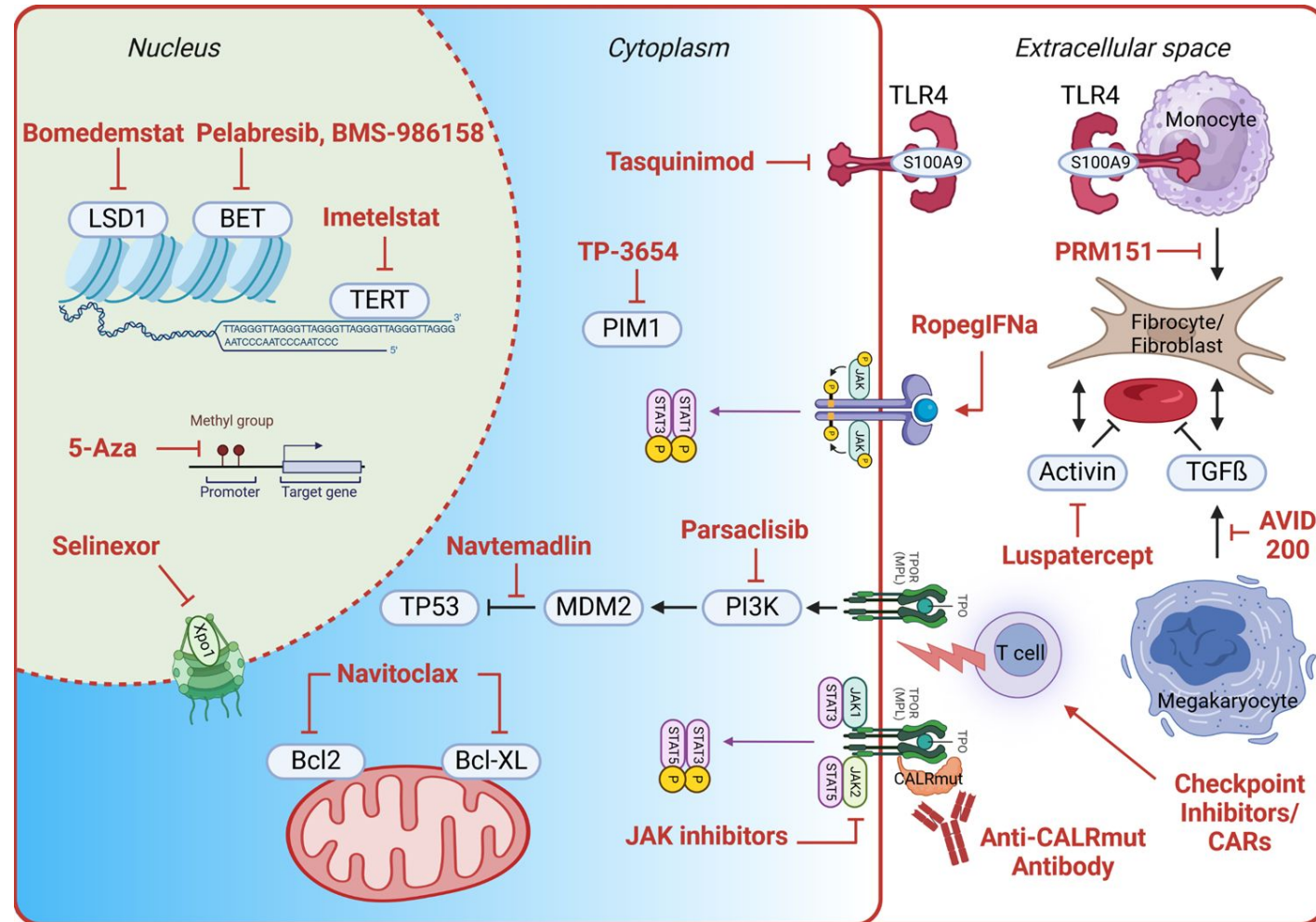
- Inflammatory cytokines
- Other soluble factors
- Cellular interactions
- Prothrombotic state
- Fibrotic stimuli
- Contribution by clonal cells (i.e. granulocytes, monocytes, platelets)
- Contribution by non-clonal cells (i.e. T and NK cells, MDSC, residual non-clonal cells)

Diagnosis

- Serum markers of inflammation
- Clonal markers („Driver and passenger mutations“)
- Diagnosis in the absence of a clonal marker (i.e. Triple-neg. MF, Triple-neg. ET, MPN-U)
- Physiologic clonal hematopoiesis

Therapy

- JAK inhibitors
- (Peg-)Interferon alpha
- Corticosteroids
- Immunomodulating drugs (IMiDs)
- Imetelstat
- Pentraxin-2
- Other drugs



Acknowledgments

Our Department, Aachen University:

Dept. of Hematology, Oncology, Hemostaseology, and SCT:

Nicolas Chatain, Marcelo Szymanski de Toledo, Julian Baumeister, Jimena Rodriguez, Milena Kalmer, Kathrin Olschok, Stefan Tillmann, Mithuoshni Arullmoli, Kim Kricheldorf, Joelle Schiffers, Manuela Klever, Mirle Schemionek, Deniz Gezer, Julia Stomper, Madeline Caduc, Rosa Cho, Rebecca Lemanyzyk, Chiara Wirths, Martina Crysandt, Gerda Silling, Tim Brümmendorf (and more)

Our collaborators:

RWTH Aachen University: Rafael Kramann, Sikander Hayat, Mirle Schemionek, Gerhard Müller-Newen, Rebekka Schneider, Martina Crysandt, Martin Zenke, Nicolas Chatain, Alexandros Sofias, Wolfgang Wagner, Hélène Gleitz, Natalia Torow, Mathias Hornef, Tim Brümmendorf, Ivan Costa, all other CRU344 members

GSG-MPN Germany: Konstanze Döhner, Martin Griesshammer, Tim Brümmendorf, Florian Heidel, Susanne Isfort, Frank Stegelmann, Haifa K. Al-Ali, Heiko Becker, Nikolas v. Bubnoff, Thomas Ernst, Thomas Fischer, Norbert Gattermann, Joachim Göthert, Madlen Jentzsch, Philipp Jost, Nicolaus Kröger, Eva Lengfelder, Heike Pahl, Markus Radsak, Andreas Reiter, Christoph Scheid, Lino Teichmann, Dominik Wolf, and other GSG-MPN members

MPN Patient Advocacy Groups: mpn-netzwerk e.V. (Germany), MPN Research Foundation (USA), MPN Advocates Network (Switzerland)

International: Alexandre Theocharides, Radek Skoda, Claire Harrison, Ruben Mesa, Jean-Jacques Kiladjian, John Mascarenhas, Naveen Pemmaraju, Raajit Rampal, Jyoti Nangalia, Tiziano Barbui, Alberto Alvarez-Larran, Simon Mendez-Ferrer, Hans Hasselbalch, Tomasz Skorski, Shannon Elf, and many others



GERMAN
STUDY
GROUP
MPN



**UNIKLINIK
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Funding Agencies:



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Thank you for your attention!



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E-Mail: skoschmieder@ukaachen.de

Personal view on MPN pathogenesis and the role of inflammation

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